



# 粗視化生体分子シミュレータ CafeMol

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# CafeMol (www.cafemol.org)



Kenzaki



- CafeMol 1.0 source & manual released
- Features are;
  - Various CG protein models
    - multiple basin model
    - accurate CG model
  - Simulating protein-at-work “switching”
- Under development DNA/RNA, lipid





# Overview of CafeMol

- General-purpose coarse-grained (CG) biomolecular modeling and simulation software
  - Protein: 1 bead / 1 amino acid
  - Nucleic acid: 3 beads (sugar, nucleotide, phosphate) / nucleotide
  - Lipid: ~3 beads / lipid
- Written by **FORTRAN90** with **MPI and Open MP**
- Large-scale simulation
  - ~"millisecond" event by K-computer
- Version 1.0 is released (only protein) (2010/12/27)



# Menu

Models

Simulation methods

Implementation

Selected applications

In-progress models & methods



# Menu

Models

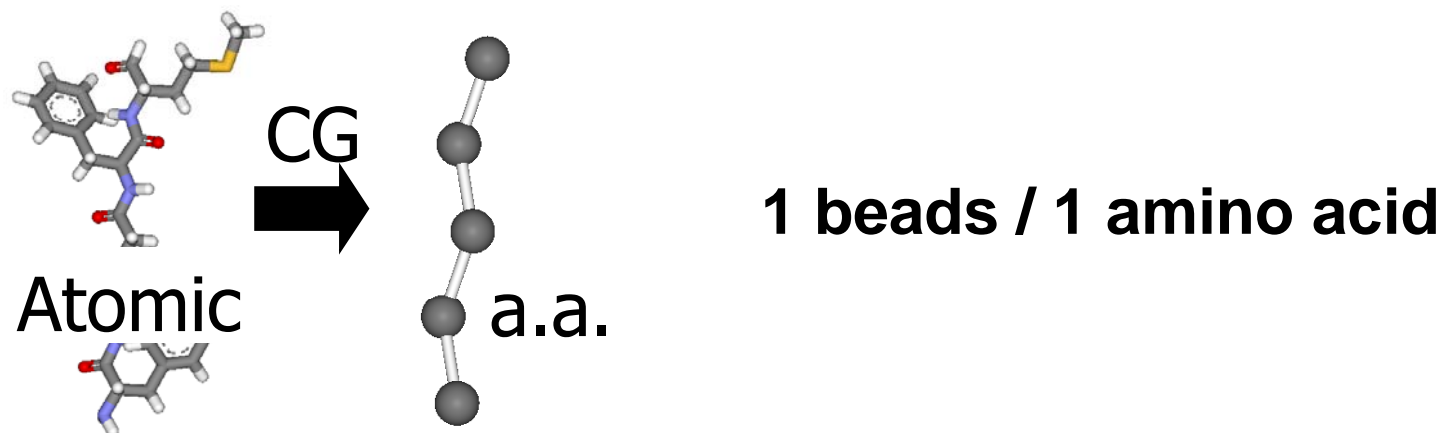
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# Models and energy functions



- A. Off-lattice Go model
- B. Atomic interaction based CG model**
- C. Multiple basin model
- D. Elastic network model
- E. Electrostatic and hydrophobic interactions
- F. Explicit and implicit ligands



# Off-lattice Go model

C. Clementi, H. Nymeyer, and J.N. Onuchic, J. Mol. Biol. (2000)

Based on the energy landscape theory  
Structure based

$$V_{protein} = V_{local} + V_{go} + V_{ex}$$

$\theta$ : bond angle  
 $\phi$ : dihedral angle  
(0 means native state)

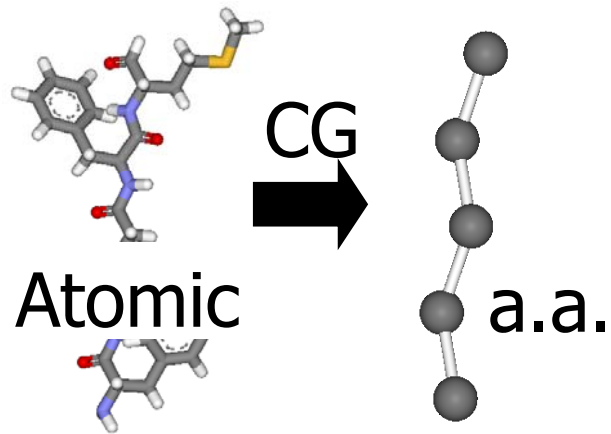
$$V_{local} = K_b \sum_i (r_{i,i+1} - r_{0i,i+1})^2 + K_\theta \sum_i (\theta_i - \theta_{0i})^2$$
$$+ K_\phi^1 \sum_i (1 - \cos(\phi_i - \phi_{0i})) + K_\phi^3 \sum_i (1 - \cos 3(\phi_i - \phi_{0i}))$$

$$V_{go} = \varepsilon_{go} \sum_{i,j}^{native} \left[ 5 \left( \frac{r_{0ij}}{r_{ij}} \right)^{12} - 6 \left( \frac{r_{0ij}}{r_{ij}} \right)^{10} \right]$$

$$V_{ex} = \varepsilon_{ex} \sum_{i,j}^{nonnative} \left( \frac{\sigma}{r_{ij}} \right)^{12}$$

$$K_b = 100\varepsilon$$
$$K_\theta = 20\varepsilon$$
$$K_\phi^1 = \varepsilon$$
$$K_\phi^3 = 0.5\varepsilon$$
$$\varepsilon_{go} = 0.18\varepsilon$$
$$\varepsilon_{ex} = \varepsilon$$
$$\sigma = 4A$$
$$\varepsilon = 1.0 \text{ kcal/mol}$$

# Atomic interaction based CG (AICG) model



$$V = \sum_i k_b^i (r^i - r_0^i)^2 + \sum_i k_a^i (\theta^i - \theta_0^i)^2 + \sum_i \{ \varepsilon_{\phi,1}^i [1 - \cos(\phi^i - \phi_0^i)] + \varepsilon_{\phi,3}^i [1 - \cos 3(\phi^i - \phi_0^i)] \} + \sum_{i>j-3}^{native} \varepsilon^{ij} [5(r_0^{ij} / r^{ij})^{12} - 6(r_0^{ij} / r^{ij})^{10}] + \sum_{i>j-3}^{non-native} \varepsilon(C / r^{ij})^{12}$$



Wenfei Li

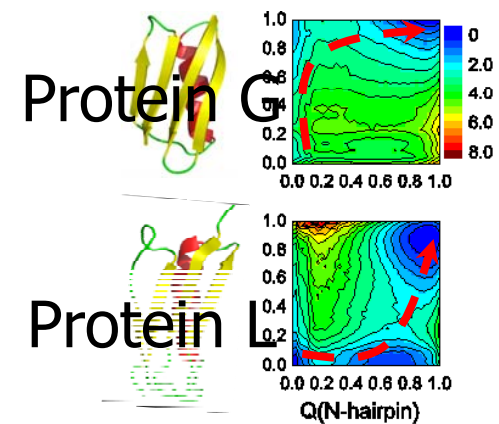
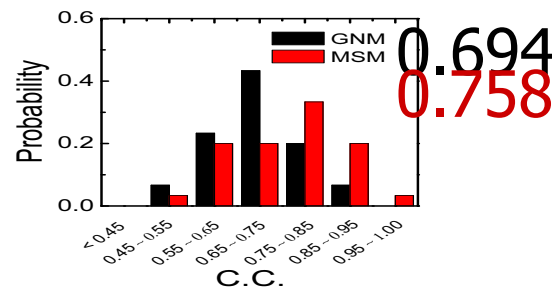
1) Contact energy  $\varepsilon_{ij}$  from pairwise all-atom (AA) energy

$$E^{IJ}(R_{IJ}) = \sum_{i \in I} \sum_{j \in J} u_{AA}(r_{ij}) \quad u_{AA}(r) = V(r) + \Delta G_{pol}^{GB}(r) + \Delta G^{SA}(r)$$

2) Coefficients fitted by AA-derived fluctuation (23 proteins)

param	$K_b$	$K_a^G$	$k_a^H$	$k_a^E$	$k_a^T$	$k_a^C$	$\varepsilon_{\phi}^G$	$\varepsilon_{\phi}^H$	$\varepsilon_{\phi}^E$	$\varepsilon_{\phi}^T$	$\varepsilon_{\phi}^C$	$\varepsilon_{nloc}$
Av.	109.94	13.40	40.0 3	17.3 2	19.35	11.7 0	0.29	1.76	1.32	0.82	0.81	0.37

Test for fluctuation, structural change, & folding





# Multiple-basin model for proteins

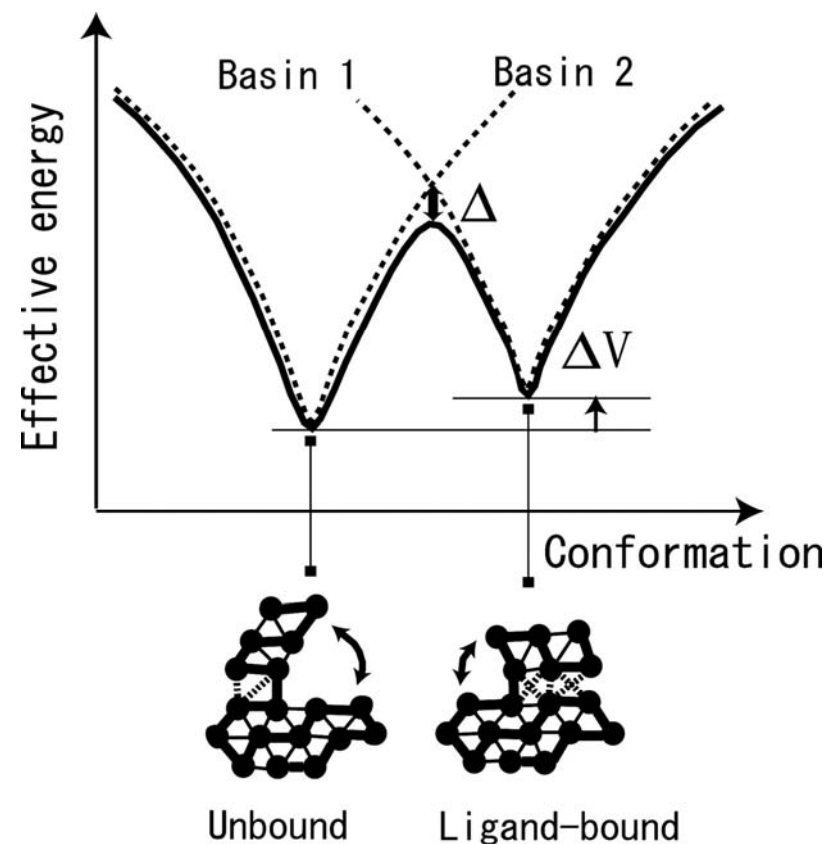
K. Okazaki, N. Koga, S. Takada, J.N. Onuchic, and P.G. Wolynes, PNAS (2006)

## Use of 2 references

$$\begin{pmatrix} V(R | R_1) & \Delta \\ \Delta & V(R | R_2) + \Delta V \end{pmatrix} \begin{pmatrix} c_1 \\ c_2 \end{pmatrix} = V_{MB} \begin{pmatrix} c_1 \\ c_2 \end{pmatrix}$$

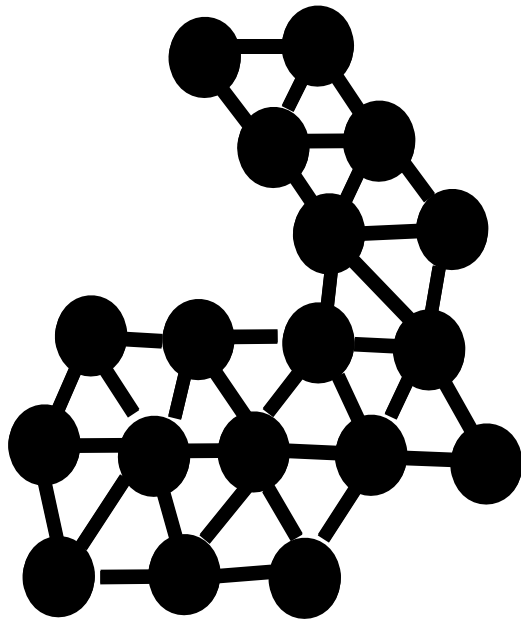
$$V_{MB} = \frac{V(R | R_1) + V(R | R_2) + \Delta V}{2} - \sqrt{\left( \frac{V(R | R_1) - V(R | R_2) - \Delta V}{2} + \Delta \right)}$$

$$\chi = \log\left(\frac{c_2}{c_1}\right)$$

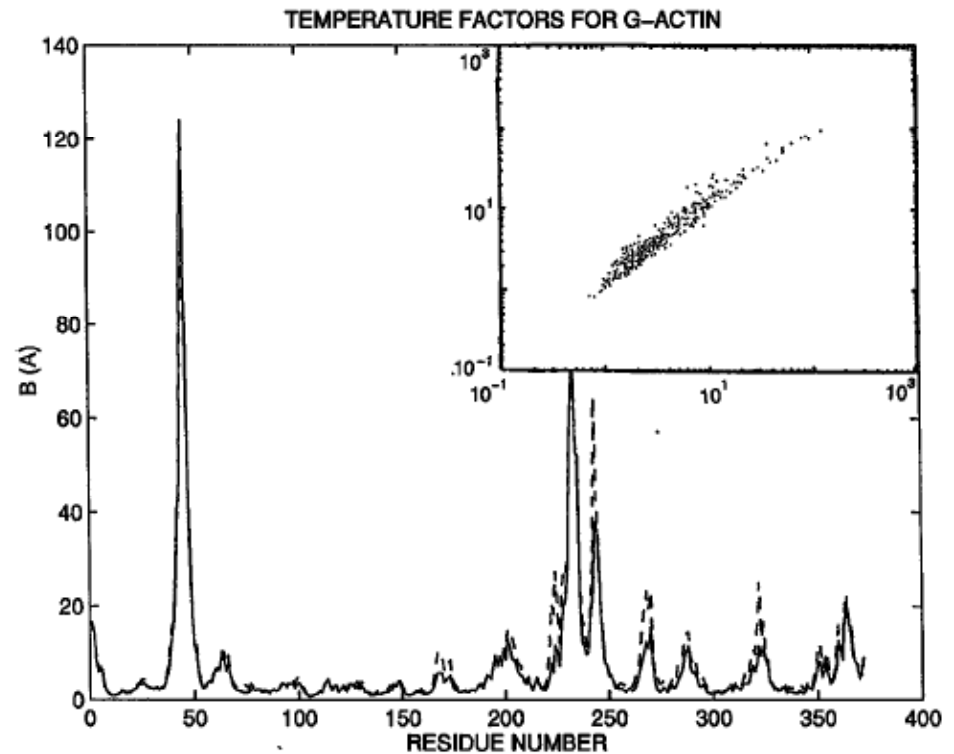


# Elastic network model

$$E = \sum_{ij, s.t. r_{ij}^0 < r_c} K (r_{ij} - r_{ij}^0)^2$$



## Atomic fluctuation reproduced by ENM (Tirion96)



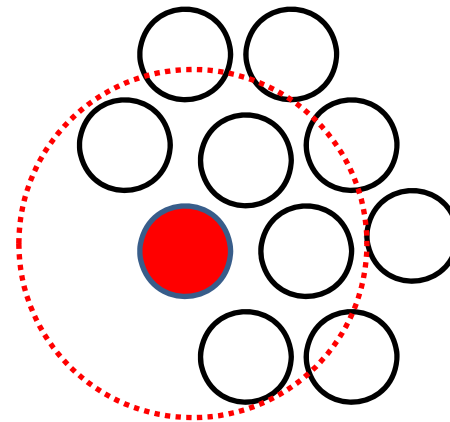
# Electrostatic and hydrophobic interactions

Debye-Huckel form for electrostatics

$$V_{\text{ele}} = \sum_{i < j}^N \frac{q_i q_j}{4\pi\epsilon_0\epsilon_k r_{ij}} e^{-r_{ij}/\kappa_D}$$

HP interactions analogous to ASA

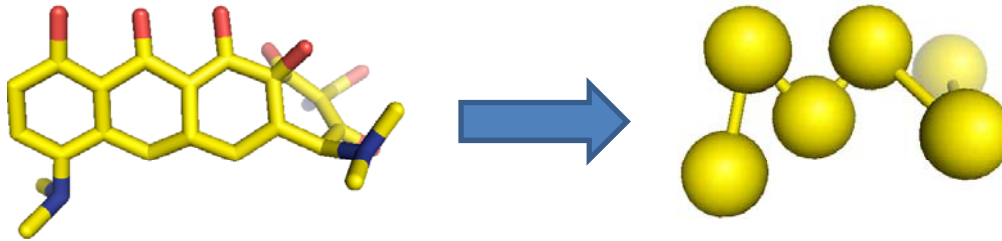
$$V_{\text{HP}} = -c_{\text{HP}} \sum_{i \in \text{HP}} \epsilon_{\text{HP},A(i)} S_{\text{HP}}(\rho_i)$$



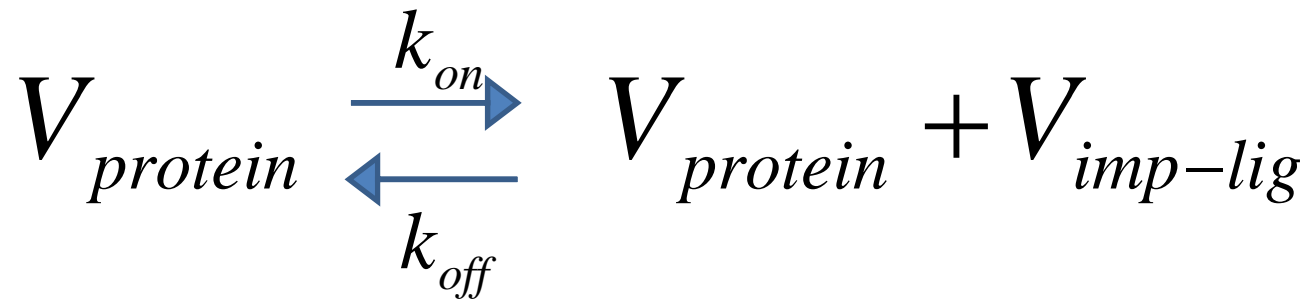
Count coordination number  
for each hydrophobic particle

# Explicit and Implicit ligands

Explicit ligand; as a rigid molecule



Implicit ligand; MD-MC scheme with ligand-mediated contact



$$V_{imp-lig} = \sum_{\text{ligand-mediated contact-pairs}} -c_{lig}\epsilon_{go} \exp \left[ -\frac{(r_{ij}/r_{0ij} - 1)^2}{2(\sigma/r_{0ij})^2} \right]$$



# Menu

Models

Simulation methods

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# Simulation method

- Dynamics
  - Newtonian dynamics with Berendsen thermostat
  - Langevin dynamics
- Time integration
  - velocity Verlet algorithm
- Run mode
  - Constant temperature simulation
  - Simulated annealing
  - Auto-search of  $T_f$
  - Replica exchange method
  - Potential “switching”



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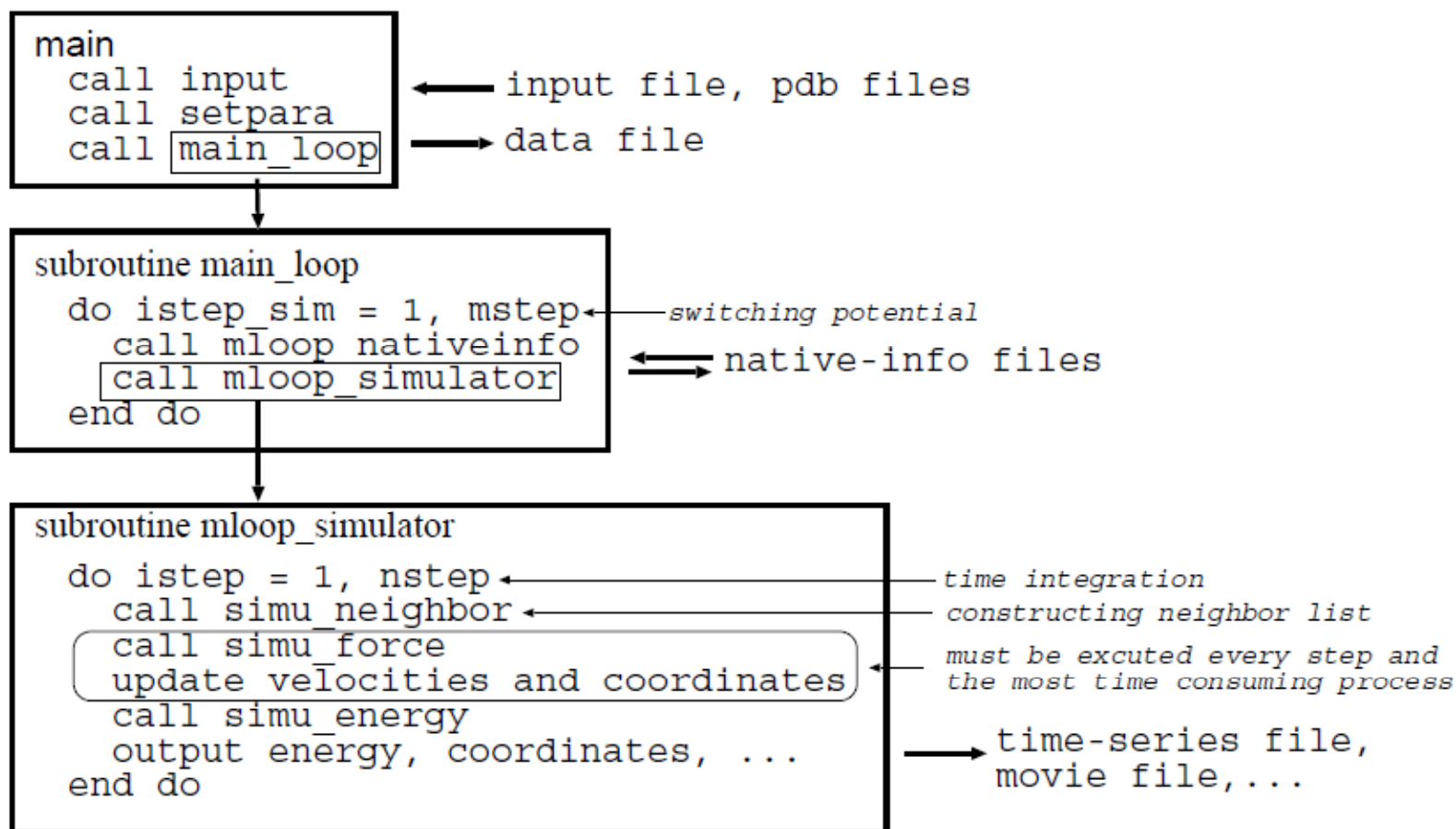
**Implementation**

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# CafeMol code



- Parallelization

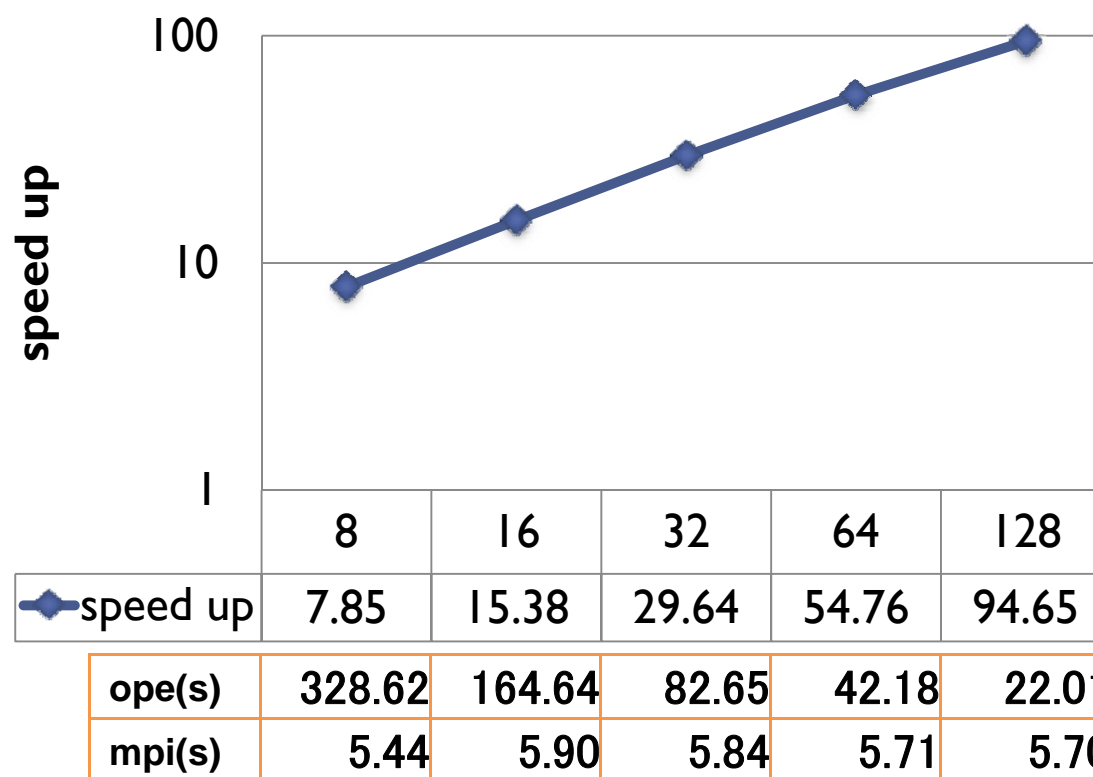
- neighboring list, force, energy  
→ hybrid(MPI+Open MP)
- replica exchange  
→ MPI(temperature/Hamiltonian REMD)





# Performance of MPI parallelization

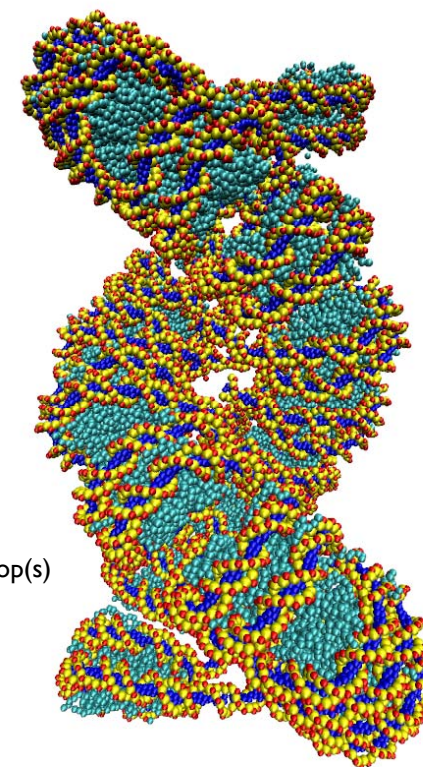
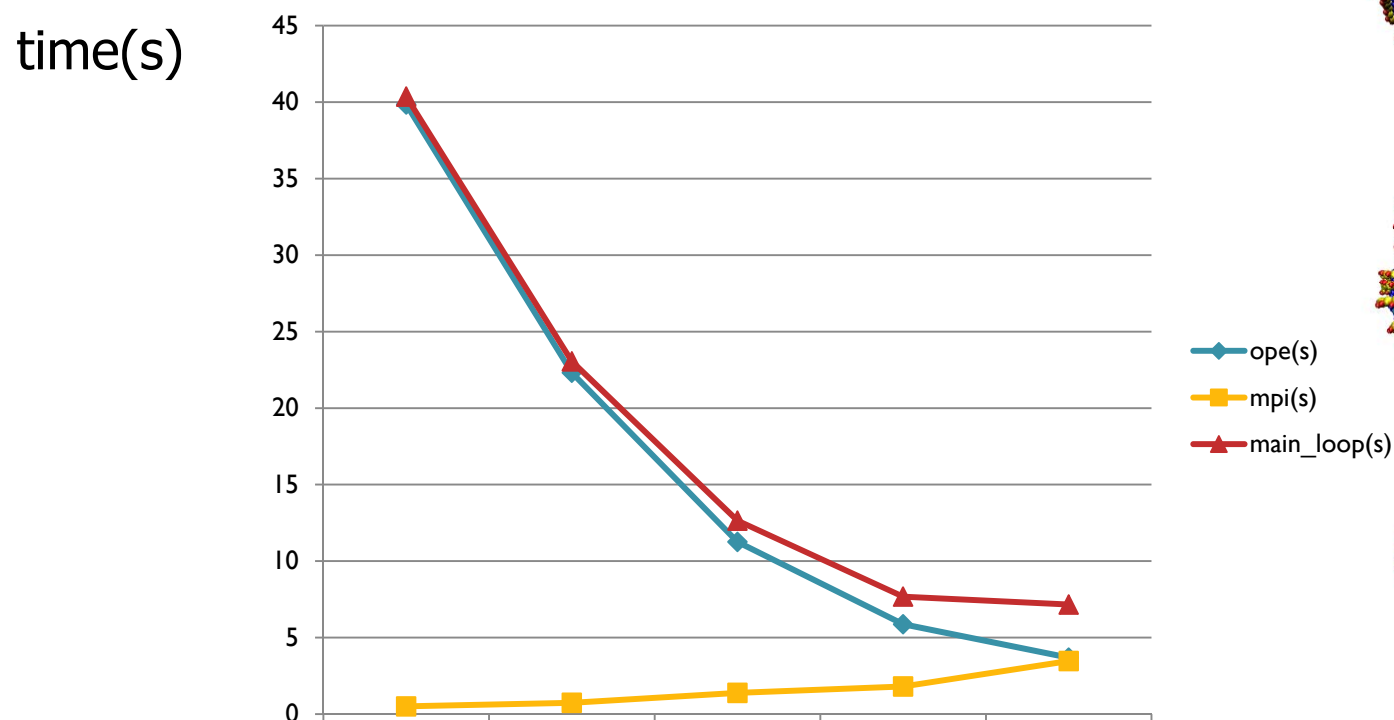
1300 base pairs DNA (7798 particles)  
BG/L at Riken



High parallelization efficiency

# Performance of hybrid parallelization (MPI and Open MP)

20 nucleosomes (35918 particles)  
RICC at Riken



OMP(force)	8	8	8	8	8
MPI(force)	1	2	4	8	15
MPI(replica)	64	64	64	64	64
speed up	1	1.75	3.19	5.26	5.64



# Menu

Models

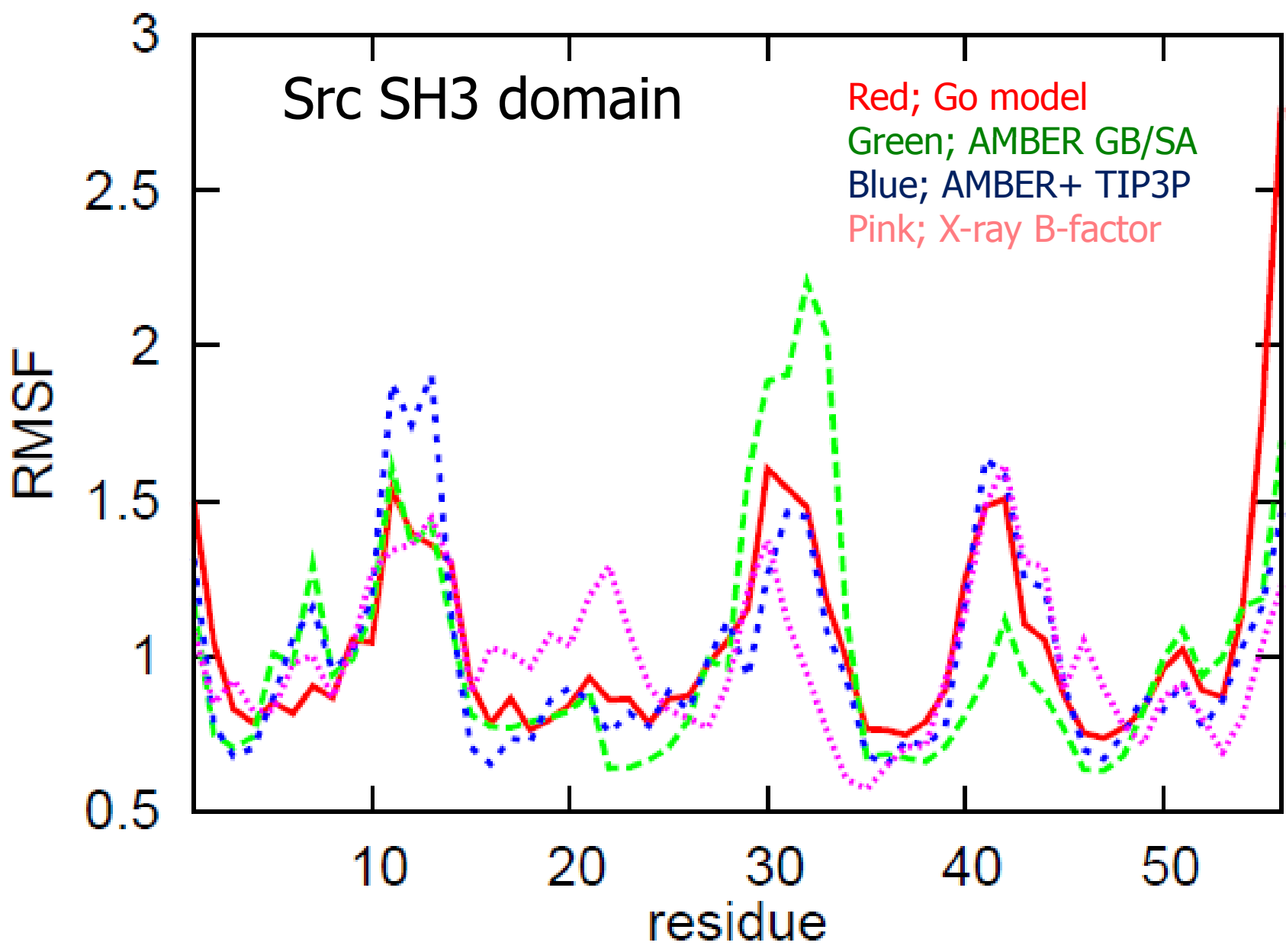
Simulation methods

Implementation

**Selected applications**

In-progress models & methods

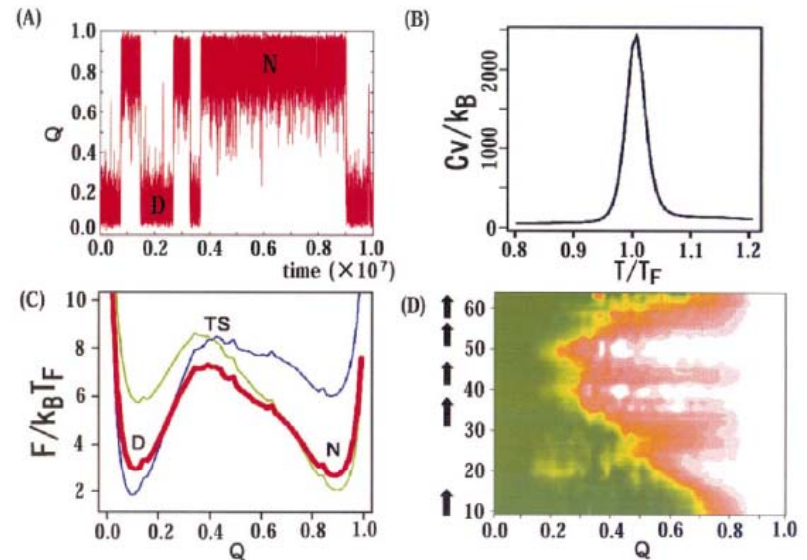
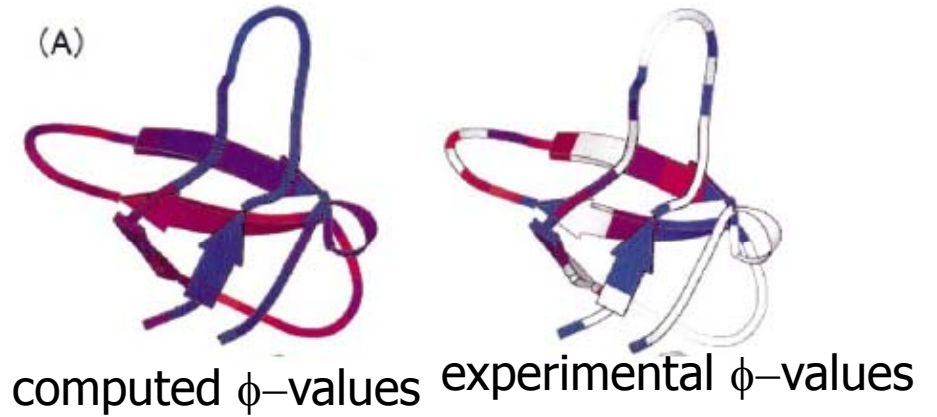
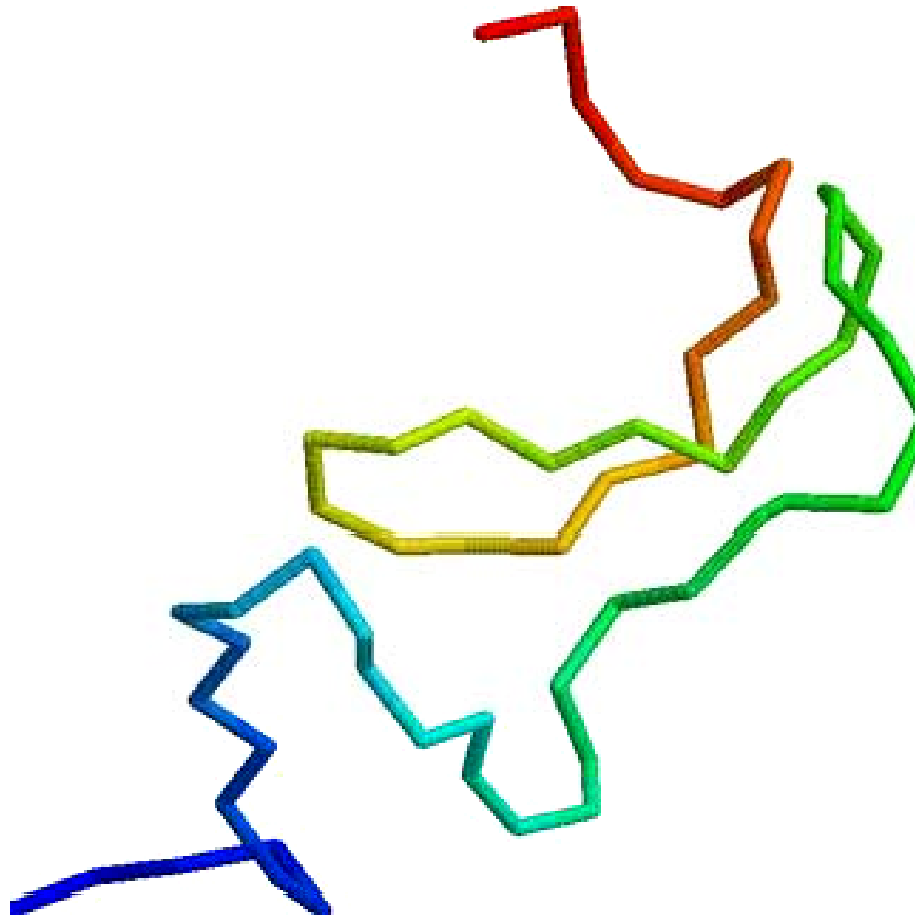
# Native fluctuation by off-lattice Go model



# Folding simulation of src SH3 domain

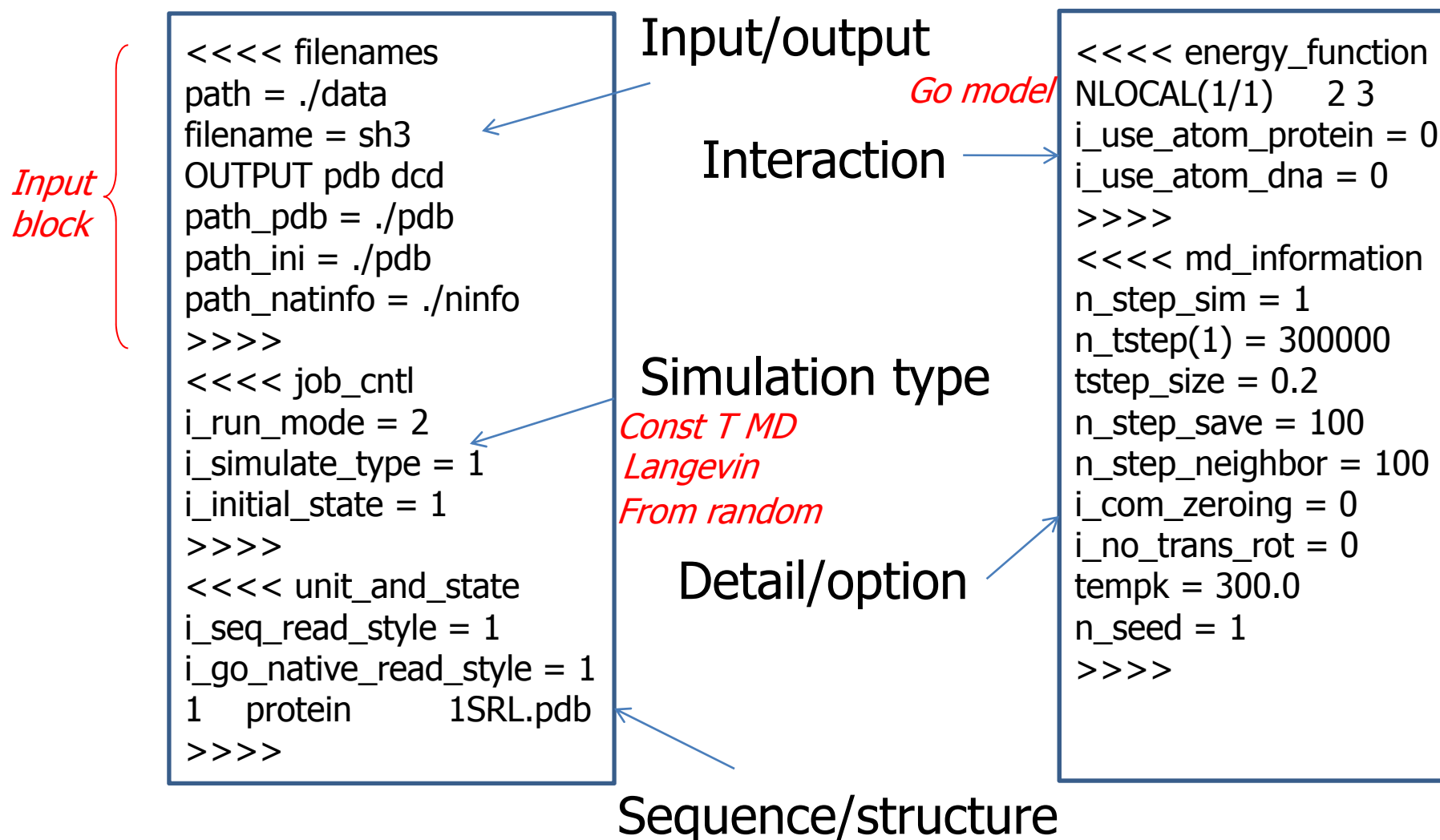


N. Koga, and S. Takada, J. Mol. Biol. (2001)





# Example of input file (folding simulation of src SH3)





# Folding temperature of src SH3 (Auto-search of Tf)

## *Bi-section method*

```
<<<< job_cntl  
i_run_mode = 4  
i_simulate_type = 1  
i_initial_state = 1  
>>>>  
<<<< searching_tf  
tempk_upper = 500.0  
tempk_lower = 100.0  
>>>>
```

```
*****  
tf_out tempk n_state d_state p_trans  
tf_out 300.000 995 5 1  
*****  
tf_out tempk n_state d_state p_trans  
tf_out 400.000 1 1000 0  
*****  
tf_out tempk n_state d_state p_trans  
tf_out 350.000 166 835 78  
*****  
tf_out tempk n_state d_state p_trans  
tf_out 325.000 953 48 19  
*****  
...  
...  
*****  
tf_out tempk n_state d_state p_trans  
tf_out 341.406 638 363 98  
*****
```



# Folding temperature of some proteins

Protein	Number of amino acid	Folding temperature(K)
albumin binding domain	53	380.4
src SH3 domain	56	342.9
protein G	56	338.2
$\alpha$ -spectrin SH3 domain	57	360.1
Sso7d	64	332.0
protein L	78	374.2
Im9	86	382.0
cytochrom B562	106	352.2





# “Switching” simulation



Native info file (*alpha\_E subunit*)

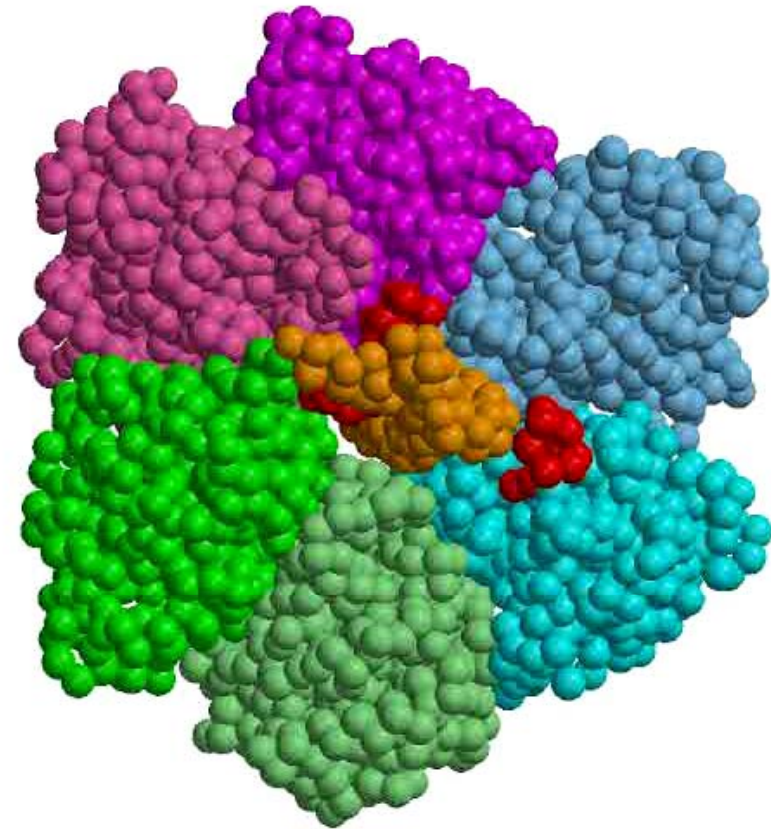
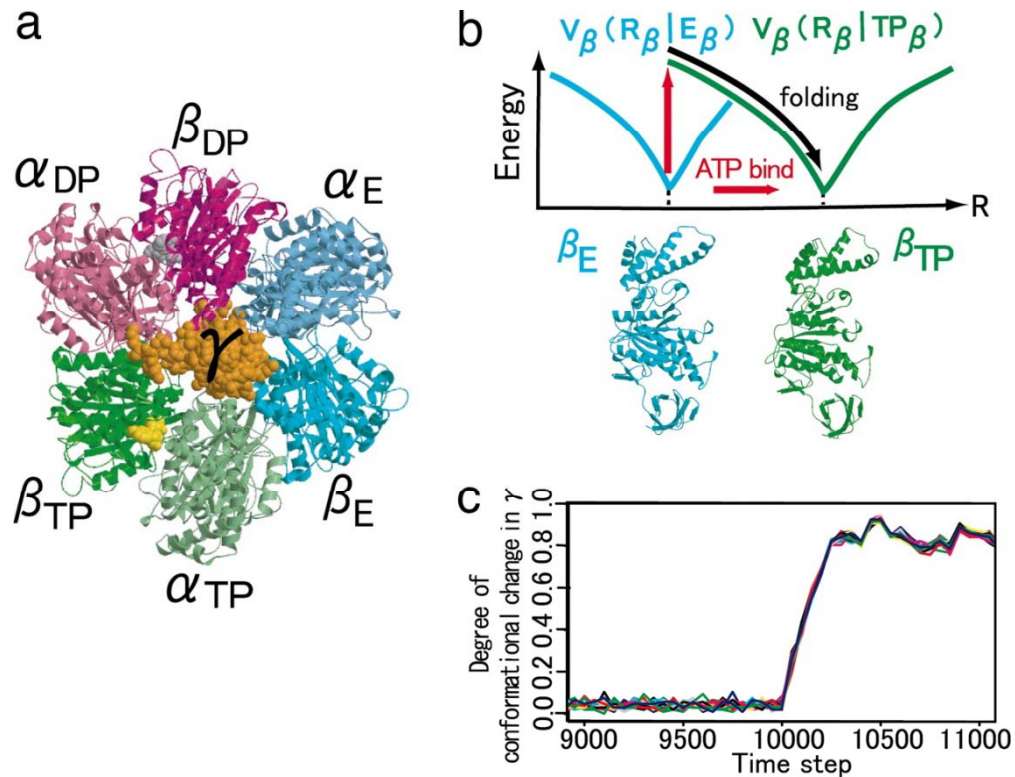
```
bond 1 1 1 1 2 1 2 3.8132 1.0000 1.0000 100.0000
...
angl 1 1 1 1 2 3 1 2 3 93.2170 1.0000 1.0000 20.0000
...
dihd 1 1 1 1 2 3 4 1 2 3 4 67.0855 1.0000 1.0000 1.0000
...
contact 1 1 1 1 5 1 5 5.9973 1.0000 1 0.3000
...
```

↑ ↑ nat- distance

↑ ↑ pair ij

# Rotation mechanism of $F_1$ -ATPase by switching Go model

N. Koga, and S. Takada, PNAS (2006)



# Conformational change by MBP



K. Okazaki, N. Koga, S. Takada, J.N. Onuchic, and P.G. Wolynes, PNAS (2006)



```
<<<< unit_and_state
i_seq_read_style = 1
i_go_native_read_style = 1
1a  protein  1GGG_2.pdb
1b  protein  1WDN_2.pdb
>>>>
<<<< energy_function
NLOCAL(1a/1a)  2 3
NLOCAL(1b/1b)  2 3
MULTIGO_SYSTEM(1a)  1a/1a
MULTIGO_SYSTEM(1b)  1b/1b
i_use_atom_protein = 0
i_use_atom_dna = 0
>>>>
<<<< multiple_go
bdemax_mgo = 100.0
baemax_mgo = 1.0
dihemax_mgo = 0.5
ENEGAP(1)(1)  0.0 -1.8
DELTA(1ab)  28.0
>>>>
```

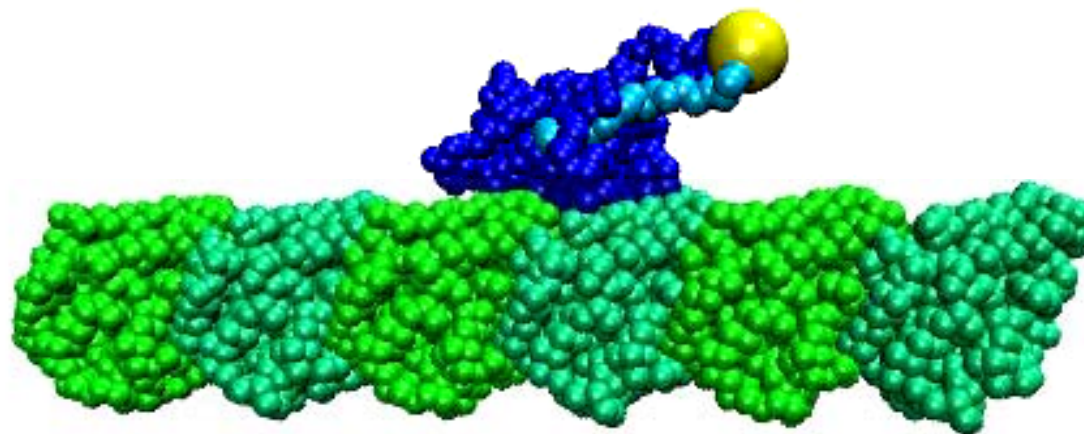


# Sliding movement of KIF1A

R. Kanada, et al unpublished data

- 1 phase: multiple-basin (T, D)
- 2 phase: go(D )
- 3 phase: multiple-basin(D, phi)
- 4 phase: go(phi)
- 5 phase: go(T)

KIF1A:blue  
tubulin:green  
carge:yellow





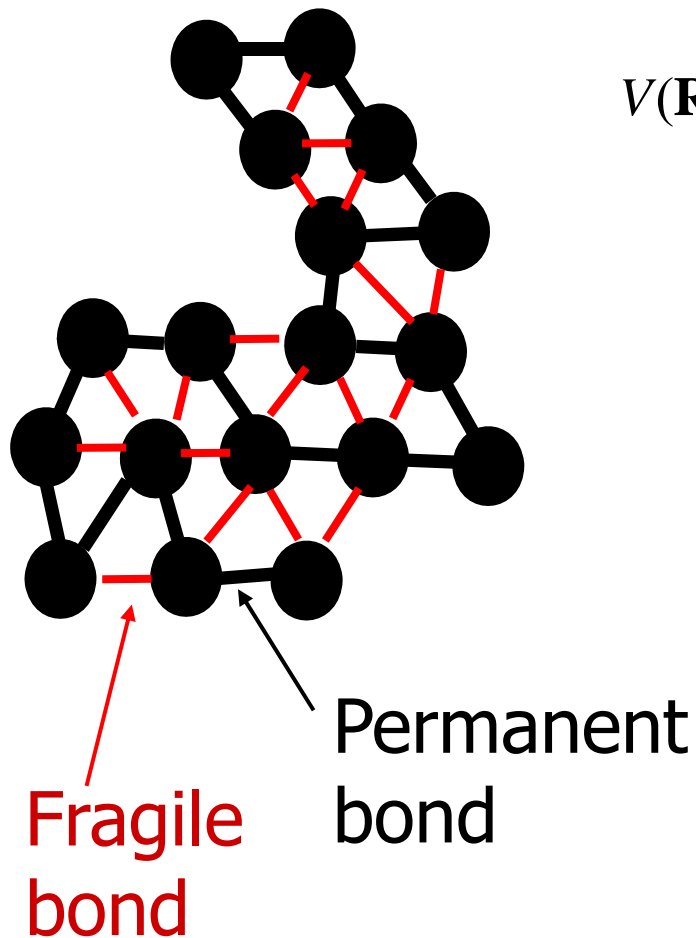
Wenfei Li

# **Frustration, specificity & nonlinearity in large-amplitude motion of allosteric proteins**

Li, Wolynes, & Takada, PNAS 2011

# Off-lattice Go model (merge of Go model & ENM)

Clementi, Nymeyer, & Onuchic 2000



$$\begin{aligned}
 V(\mathbf{R}|\mathbf{R}_0) = & \sum_{\text{bonds}} K_r (b_i - b_{i_0})^2 + \sum_{\text{angles}} K_\theta (\theta_i - \theta_{i_0})^2 + \\
 & \sum_{\text{dihedral}} \{K_\phi^1 [1 - \cos(\phi_i - \phi_{i_0})] + K_\phi^3 [1 - \cos(3(\phi_i - \phi_{i_0}))]\} + \\
 & \sum_{i < j-3}^{\text{natcontact}} \varepsilon_1 \left[ 5 \left( \frac{r_{0ij}}{r_{ij}} \right)^{12} - 6 \left( \frac{r_{0ij}}{r_{ij}} \right)^{10} \right] + \sum_{i < j-3}^{\text{non-nat}} \varepsilon_2 \left( \frac{D}{r_{ij}} \right)^{12}
 \end{aligned}$$

Subscript 0 means  
the value at native

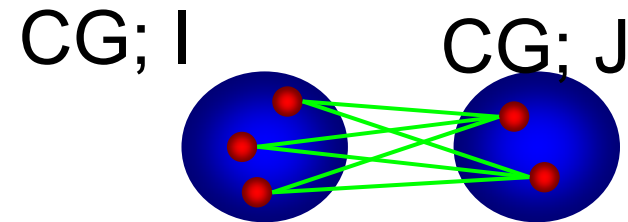
$$\begin{aligned}
 K_r = 100e \quad K_\theta^1 = 20e \quad K_\phi^3 = 1.0e \quad K_\phi = 0.5e \\
 \varepsilon_1 = \varepsilon_2 = 0.17e \quad D = 0.45(\text{\AA})
 \end{aligned}$$

# Atomic interactions in allosteric proteins: Energy decomposition

Analysis of residue-pairwise energy from atomic force field

$$E^{IJ}(R_{IJ}) = \sum_{i \in I} \sum_{j \in J} u_{AA,ij}(r)$$

*Gohlke et al., JMB, 2003, 330, 891*



$E^{IJ}$ : coarse grained contact

energy  
 $u_{AA,ij}$ : All atom energy between atom

pair  $(i, j)$

$$U_{AA}(r) = V(r) + \Delta G_{pol}^{GB}(r) + \underline{\Delta G^{SA}(r)}$$

Using LCPO (pair wise)

AA energy include vacuum part and solvation part, by **AMBER99SB** force field.

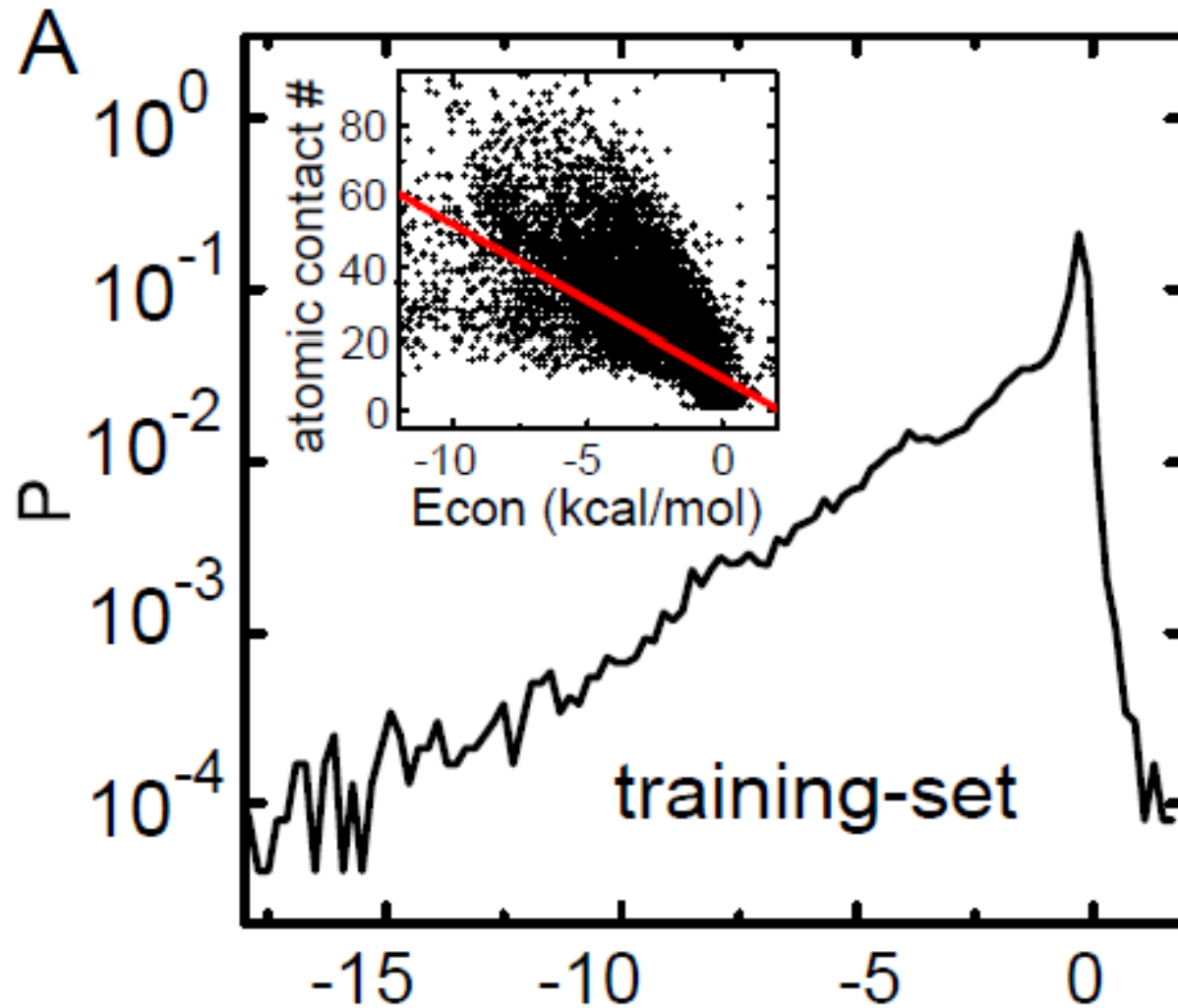
*Case et al. AMBER10, 2008*

*Weiser et al, JCC, 1999, 20,217*

# Contact energies in single-domain proteins

## Energy decomposition

Exponential law!!

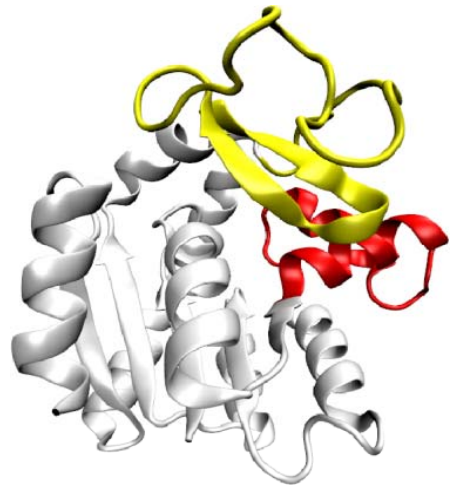




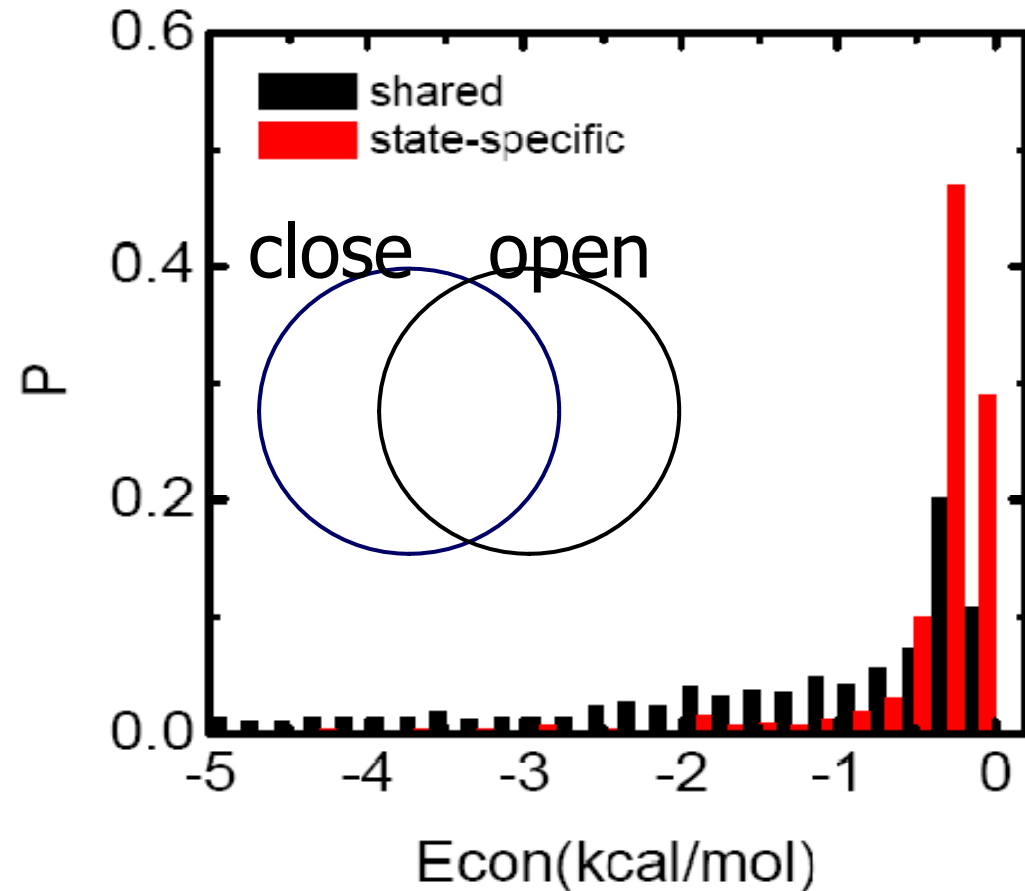
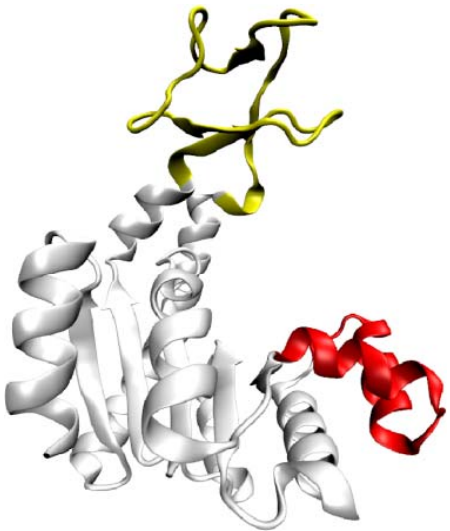
# Contact energies in allosteric proteins: Energy decomposition

## Adenylate kinase

1ake  
close



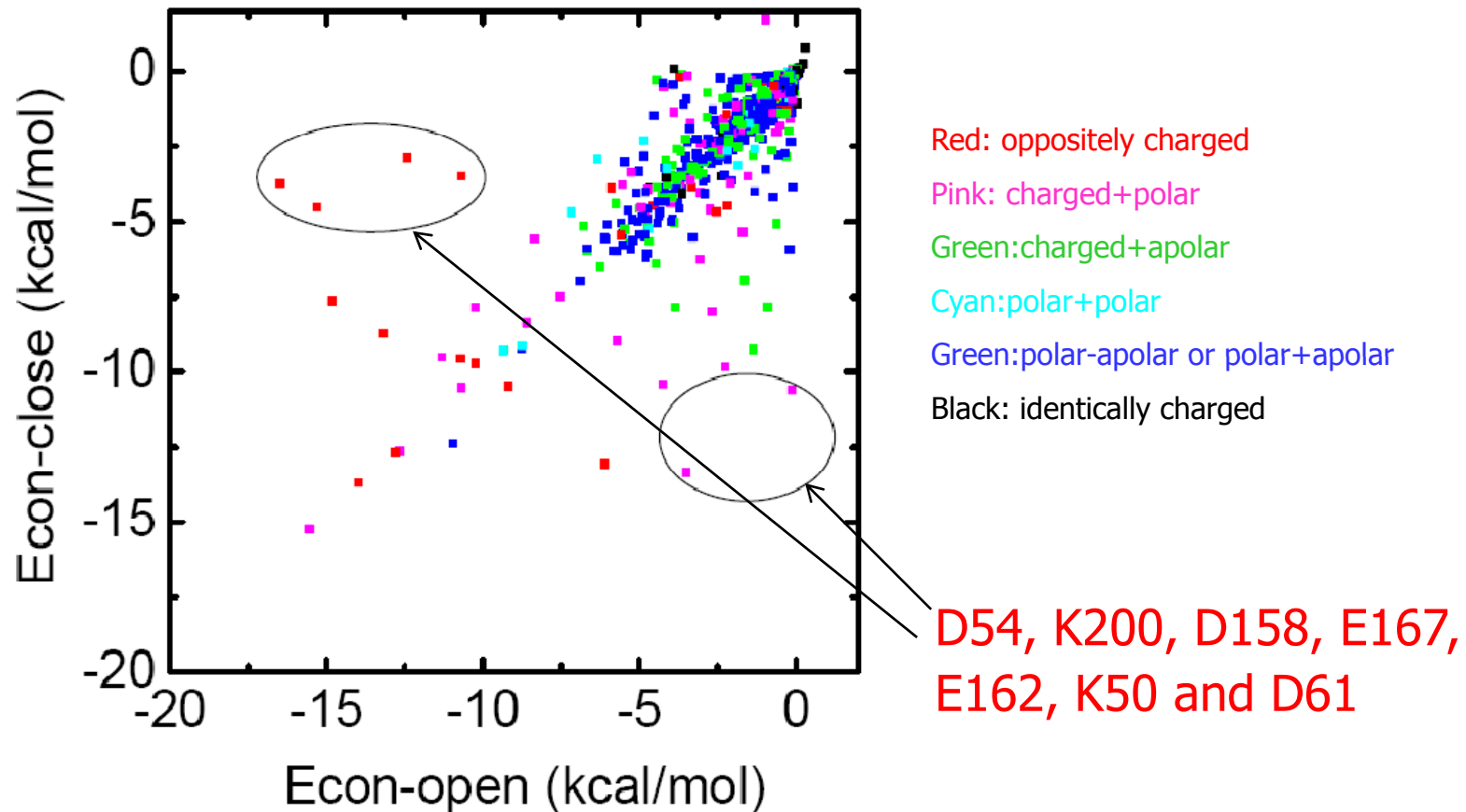
4ake  
open



- Diverse
- Diff bet. classes

# Atomic interactions in allosteric proteins: Energy decomposition

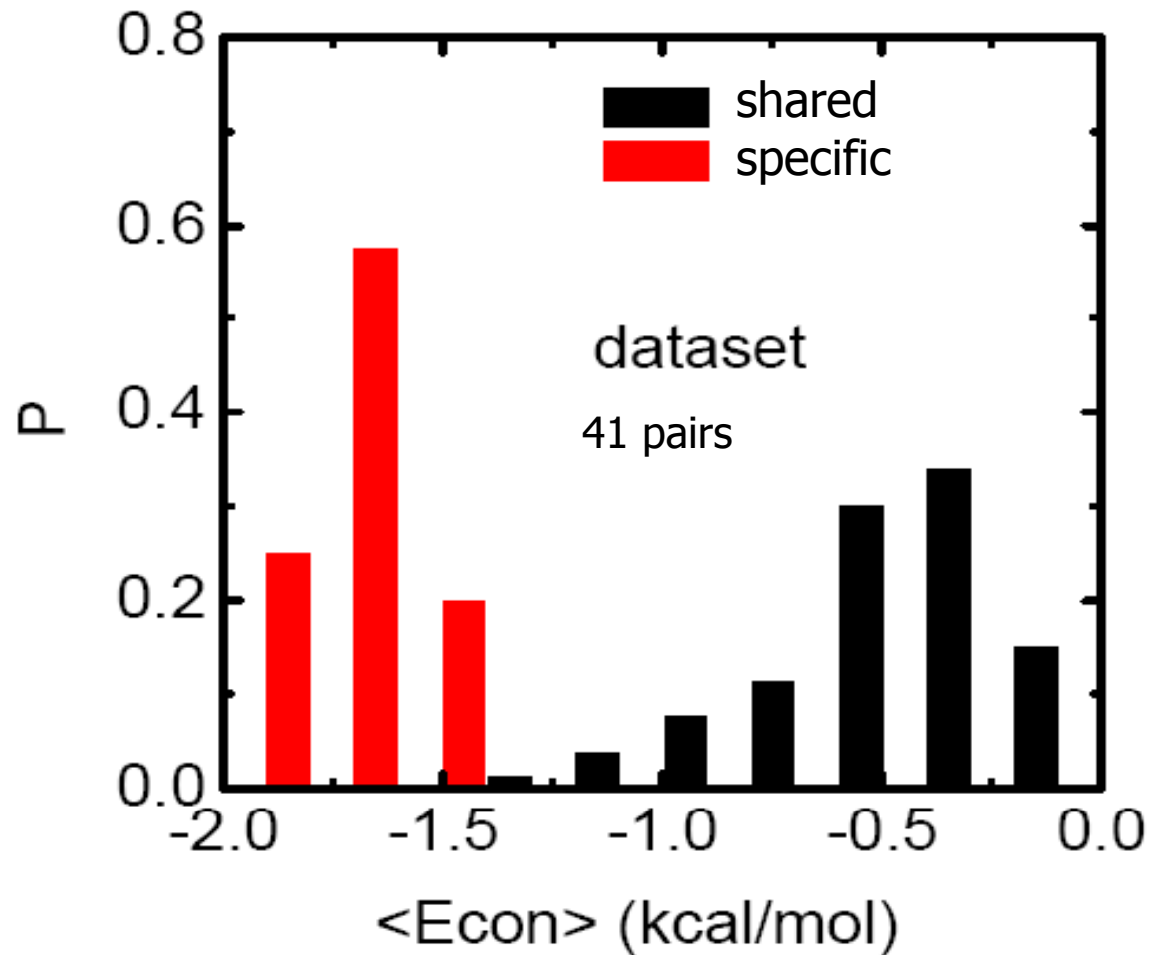
Adenylate kinase



# Atomic interactions in allosteric proteins: Energy decomposition

41 allosteric proteins in pdb

Comp of class-average for each protein



# Atomic interaction based CG (AICG) model

## Deriving parameters

$$\begin{aligned} V(R|R^0) = & \sum_i k_b^i (r^i - r_0^i)^2 \\ & + \sum_i k_a^i (\theta^i - \theta_0^i)^2 \\ & + \sum_i \{ \varepsilon_\phi^i [1 - \cos(\phi^i - \phi_0^i)] \\ & + \varepsilon_\phi^i [1 - \cos 3(\phi^i - \phi_0^i)] / 2 \} \\ & + \sum_{i>j-3}^{native} \varepsilon^{ij} [5(r_0^{ij} / r^{ij})^{12} - 6(r_0^{ij} / r^{ij})^{10}] \\ & + \sum_{i>j-3}^{non-native} \varepsilon (C / r^{ij})^{12} \end{aligned}$$

Multiscale strategies:

- ❖ Energy decomposition → relative weight of contact energies
- ❖ Match rmsf bet. AA and CG → scale local & tertiary interactions

*Chu and Voth, 2006*

*Li et al., 2010,*

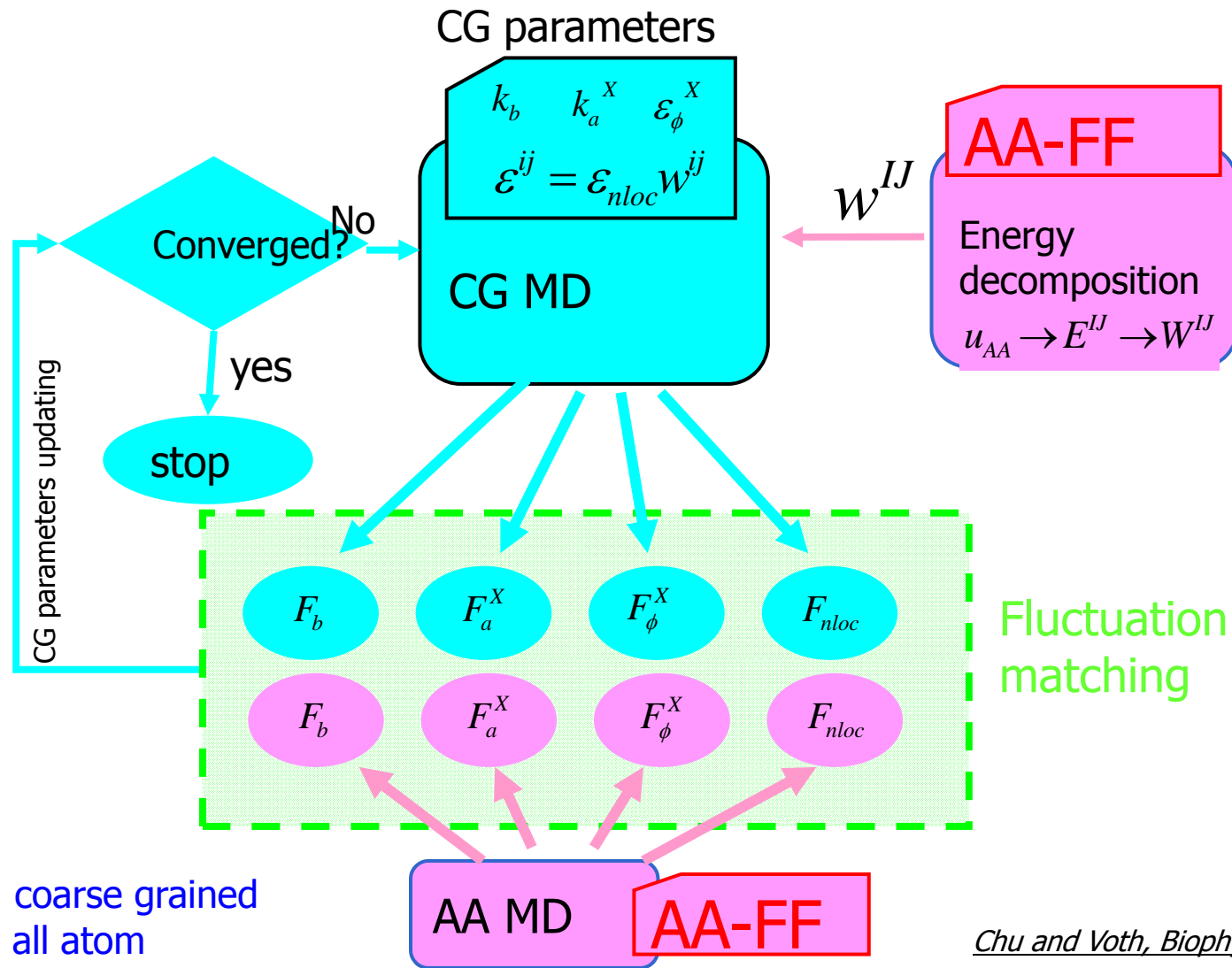
*See also*

*Trylska et al*

*Gohlke et al., 2003,*

For 23 training proteins

--- Flow chart



CG: coarse grained  
 AA: all atom  
 FF: force field

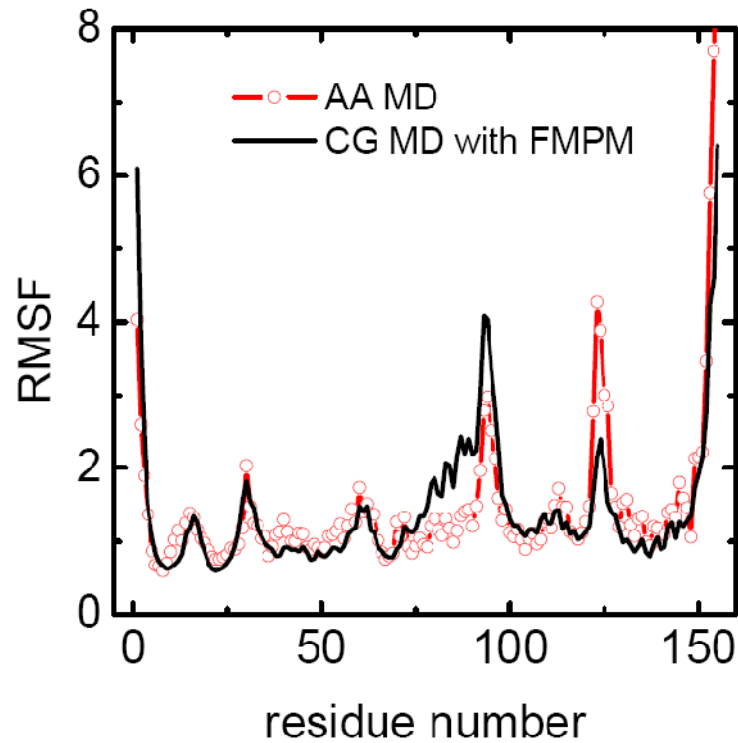
*Chu and Voth, Biophys. J., 2006,90,1572*

$F$ : mean square fluctuations

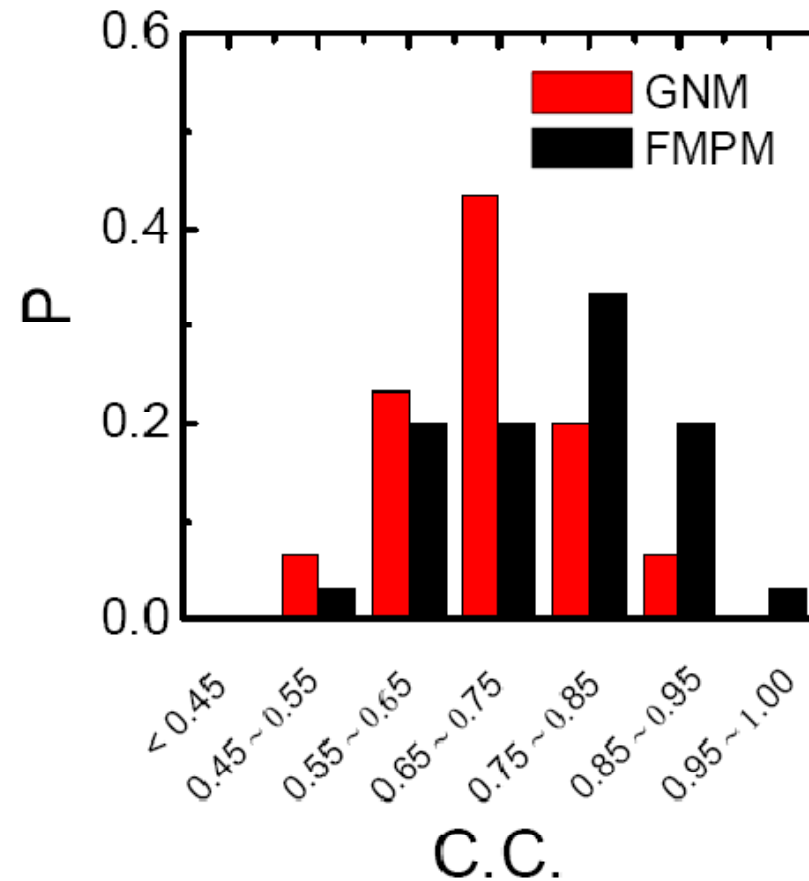
# AICG model for mean fluctuations

Near native fluctuation (RMSFs)

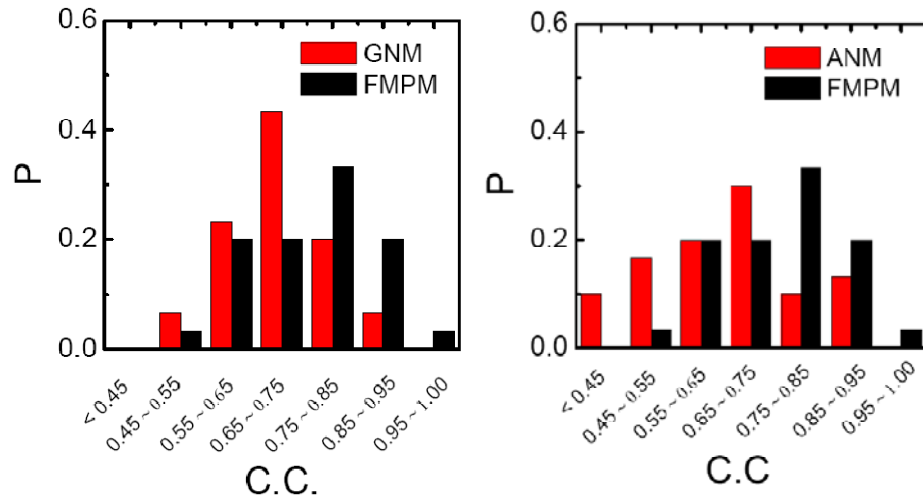
## Comp against AA MD



## Stat for 30 proteins



# AICG model for mean fluctuations



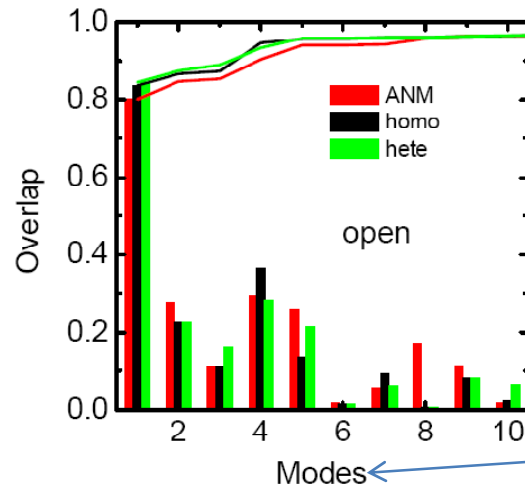
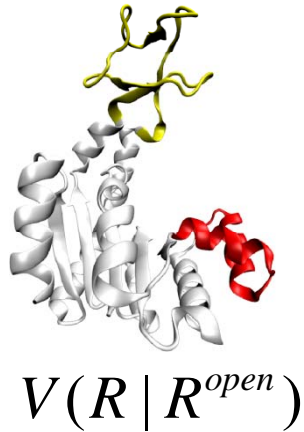
30 proteins

Fujitsuka et al, 2006, *Proteins*  
Yang et al, 2007, *Structure*

Table 3. Average correlation coefficients (C.C.) and standard errors (S.E.) between the rmsfs derived by AA model and by different CG models based on the proteins of testing set.

	ENM		AICG model		
Models	GNM	ANM	hete	homo	hete-nloc
C.C.	0.694	0.648	<b>0.758</b>	0.722	0.738
S.E.	0.018	0.031	0.021	0.027	0.031
<p><b>hete:</b> heterogeneous model.  <b>homo:</b> homogeneous model.  <b>hete-nloc:</b> only the nonlocal interactions are heterogeneous.</p>					

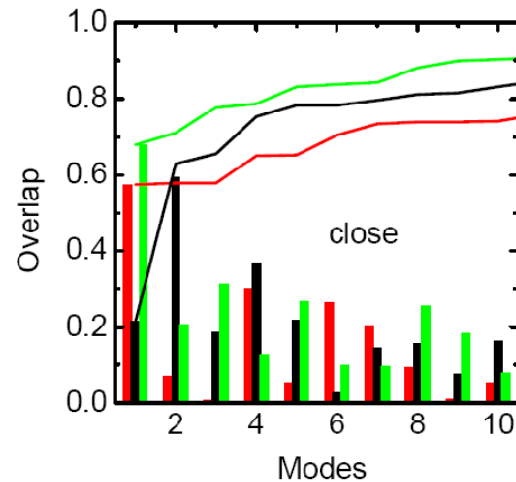
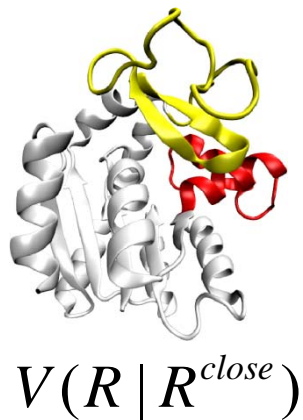
# AICG model for predicting structural change adenylate kinase



$$\vec{d} = (\vec{R}^o - \vec{R}^c) / |\vec{R}^o - \vec{R}^c|$$

$$overlap(i) = \vec{v}^i \cdot \vec{d}$$

PCA for Go model,  
NM for ANM





# AICG model for predicting structural change

## Correlation w. structure change

### 41 struct pairs of allosteric proteins

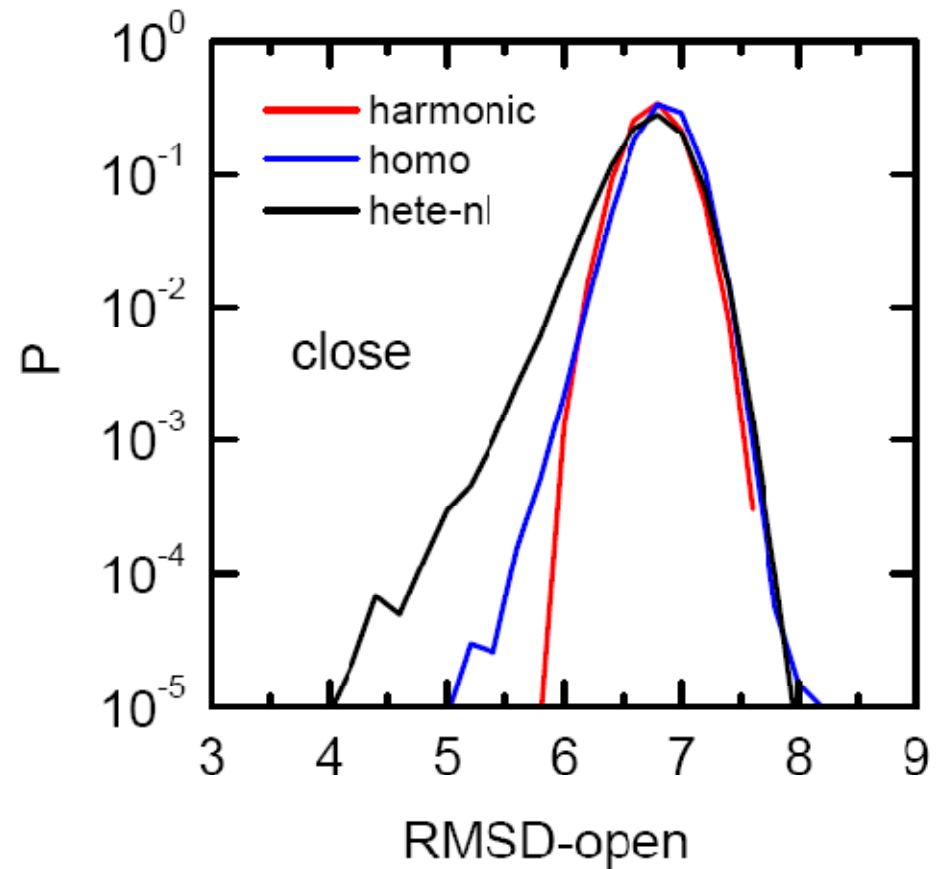
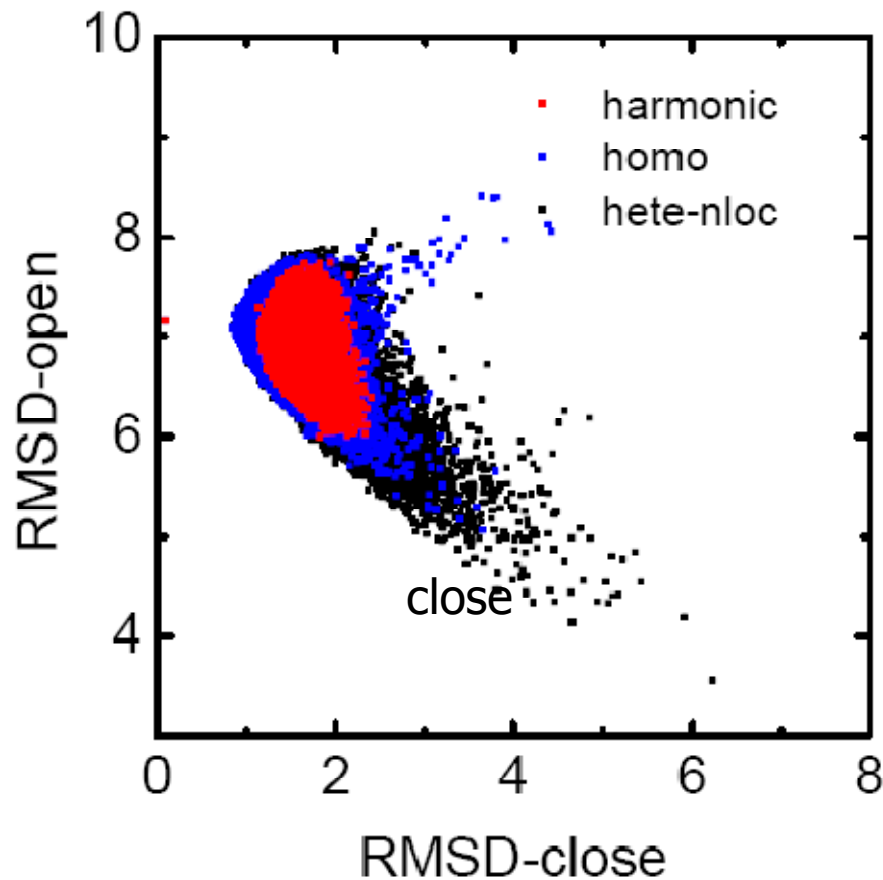
Gerstein, NAR1998

		ENM	New model		
Models		ANM	hete	homo	hete-nloc
open→close $V(R   R^{open})$	M.O.	0.480 (0.037)	0.540 (0.037)	0.511 (0.038)	0.518 (0.040)
	C.O.	0.600 (0.040)	0.657 (0.039)	0.628 (0.040)	0.626 (0.042)
close→open $V(R   R^{close})$	M.O.	0.421 (0.032)	0.517 (0.036)	0.475 (0.036)	0.512 (0.038)
	C.O.	<b>0.556</b> (0.037)	<b>0.638</b> (0.038)	0.608 (0.038)	0.631 (0.039)

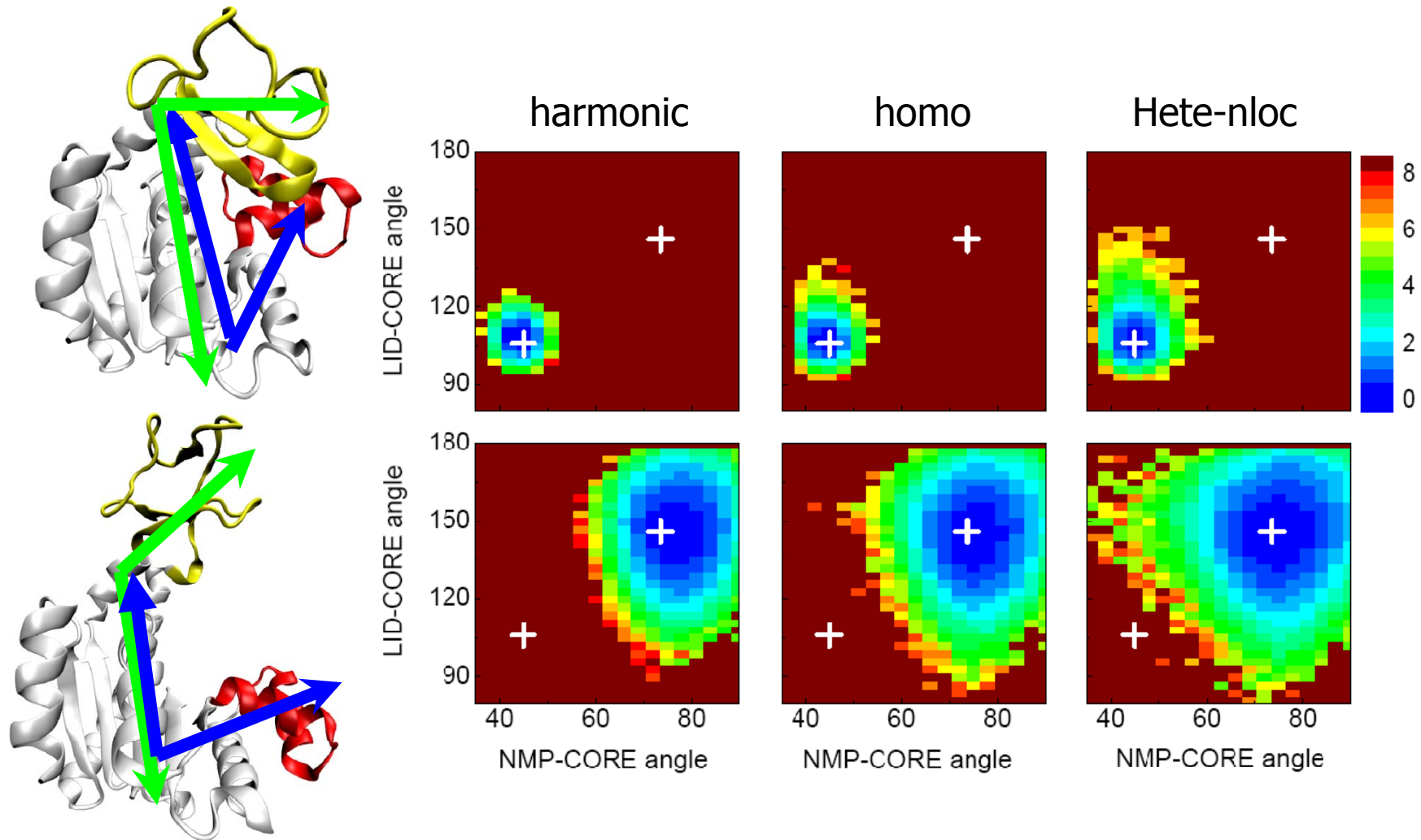
Both the interaction heterogeneity and anharmonicity are important for predicting the conformational change.

# Large-amplitude fluctuation adenylate kinase

$V(R | R^{close})$  Fluctuation in close state

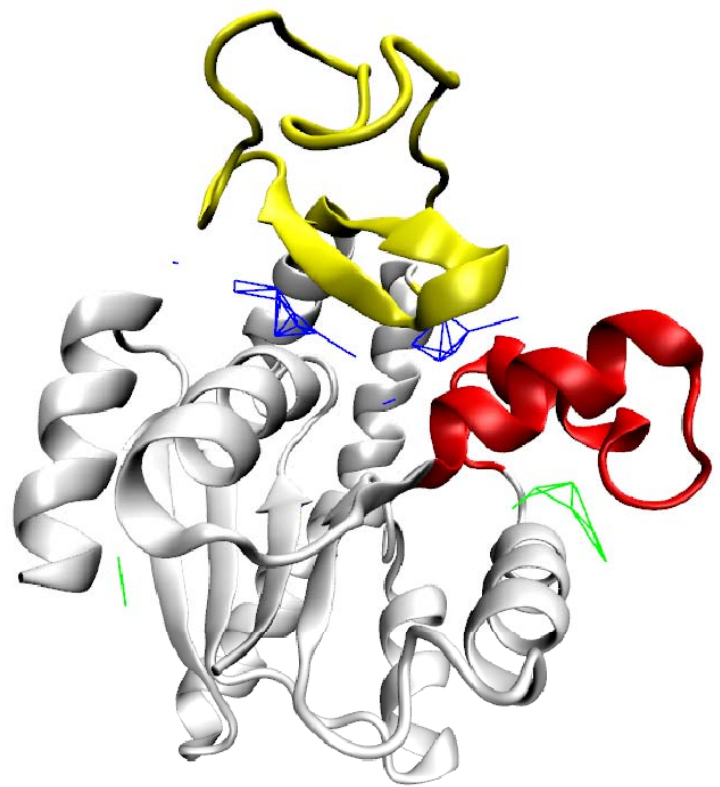
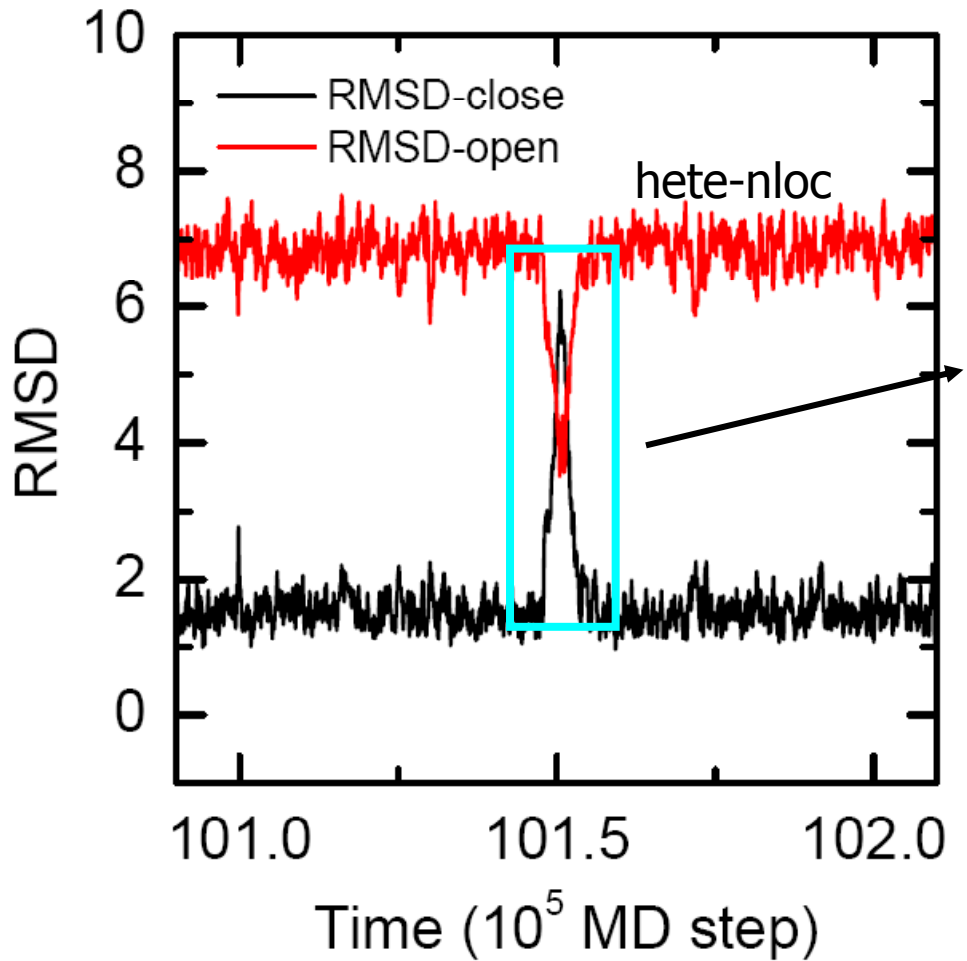


# Large-amplitude fluctuation in AKE



LID domain has higher mobility than the NMP domain.  
Interaction heterogeneity enhances the collective motions.

# Large-amplitude fluctuation in AKE



Very rare event in a long trajectory



# Menu

Models

Simulation methods

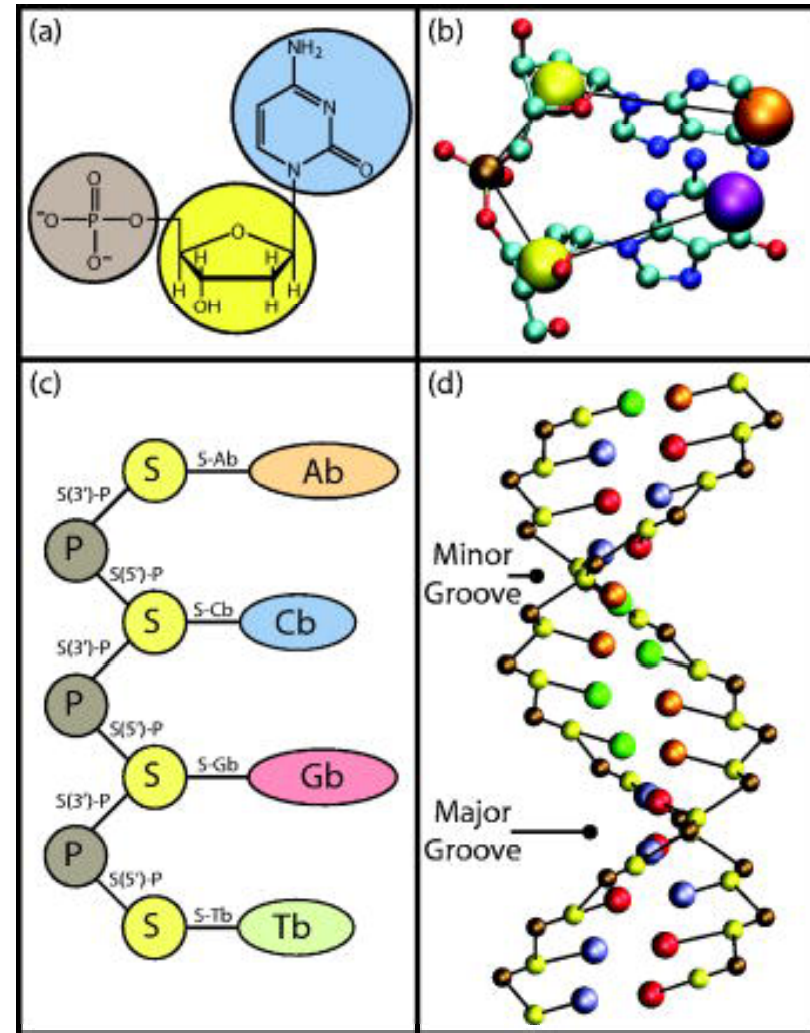
Implementation

Selected applications

**In-progress models & methods**

# CG DNA model

- Three interaction sites
  - Phosphate
  - Sugar
  - Base
- Reproduce various DNA behavior
  - Salt-dependent melting
  - Bubble formation
  - Mechanical properties



# 3SPN.1 force field

E.J. Sambrisiki, D.C. Schwartz, and J.J. de Pablo, Knotts, Biophys J. (2009)

$$V_{dna} = V_{local} + V_{stack} + V_{bp} + V_{ex} + V_{qq} + V_{solv}$$

$$V_{local} = K_{b1} \sum_i (r_{i,i+1} - r_{0i,i+1})^2 + K_{b2} \sum_i (r_{i,i+1} - r_{0i,i+1})^4$$

$$+ K_{\theta} \sum_i (\theta_i - \theta_{0i})^2 + K_{\phi} \sum_i (1 - \cos(\phi_i - \phi_{0i}))$$

$$V_{stack} = 4\epsilon_1 \sum_{i,j}^{N_{st}} \left[ \left( \frac{\sigma_{0ij}}{r_{ij}} \right)^{12} - \left( \frac{\sigma_{0ij}}{r_{ij}} \right)^6 \right]$$

$$V_{bp} = \sum_{i,j}^{N_{bp}} 4\epsilon_{bpi} \left[ 5 \left( \frac{r_{0ij}}{r_{ij}} \right)^{12} - 6 \left( \frac{r_{0ij}}{r_{ij}} \right)^{10} \right]$$

$$V_{ex} = 4\epsilon_1 \sum_{i,j}^{N_{ex}} \left[ \left( \frac{\sigma_0}{r_{ij}} \right)^{12} - \left( \frac{\sigma_0}{r_{ij}} \right)^6 \right] + \epsilon_1 \text{ (if } r_{ij} < d_{cut}),$$

$$= 0 \text{ (if } r_{ij} > d_{cut})$$

$\theta$ : bond angle  
 $\phi$ : dihedral angle  
 (0 means B-type DNA)

Go type interaction

$$\begin{aligned} K_{b1} &= 1\epsilon \\ K_{b2} &= 100\epsilon \\ K_{\theta} &= 1400\epsilon \\ K_{\phi} &= 28\epsilon \\ \epsilon_{bpGC} &= 2.532\epsilon \\ \epsilon_{bpAT} &= 2.0\epsilon \\ \epsilon &= 0.1839 \text{ kcal/mol} \end{aligned}$$



# 3SPN.1 force field (electrostatic and solvation interaction)

$$V_{qq} = \sum_{i,j}^N \left( \frac{q_i q_j}{4\pi\epsilon_0\epsilon(T,C)r_{ij}} \right) e^{-r_{ij}/\kappa D} \quad \leftarrow \text{Debye-Huckel theory}$$

$$\epsilon(T,C) = \epsilon(T)a(C)$$

$$\epsilon(T) = 249.4 - 0.788T / K + 7.20 \times 10^{-4} (T / K)^2$$

$$a(C) = 1.000 - 0.2551C / M$$

$$+ 5.151 \times 10^{-2} (C / M)^2 - 6.889 \times 10^{-3} (C / M)^3$$

$$V_{solv} = \sum_{i<j}^{N_{solv}} \epsilon_s \left[ 1 - e^{-a(r_{ij}-r_s)} \right]^2 - \epsilon_s$$

$$\epsilon_s = \epsilon_N A_I$$

$$e_N = e_0 (1 - [1.40418 - 0.268231 N_{nt}]^{-1})$$

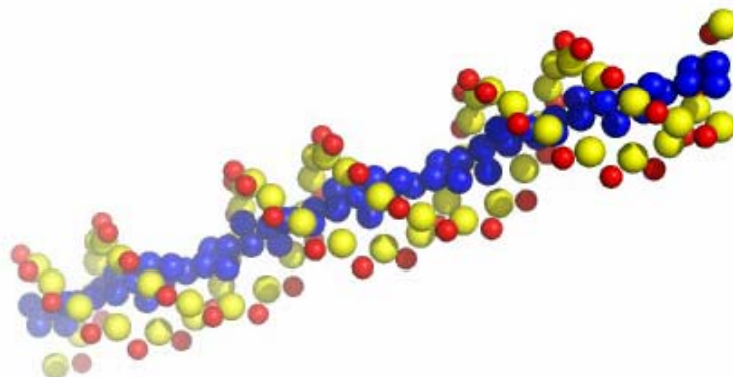
$$A_I = 0.474876 (1 + \{0.148378 + 10.9553 [Na^+]\}^{-1})$$

$$\begin{aligned} \alpha^{-1} &= 5.333\text{\AA} \\ r_s &= 13.38\text{\AA} \\ \epsilon_0 &= 0.504982\epsilon \end{aligned}$$



# DNA duplex

- 30 bp oligomer of DNA
- Langevin dynamics (300K)
- $[Na^+] = 69mM$



```

<<<< unit_and_state
i_seq_read_style = 2
i_go_native_read_style = 3
1-2 dna sequence
>>>>
<<<< energy_function
NLOCAL(1-2/1-2) 7 11
i_use_atom_protein = 0
i_use_atom_dna = 0
>>>>
<<<< electrostatic
cutoff_ele = 20.0
ionic_strength = 0.069
diele_water = 78.0
>>>>
<<<< in_box
xbox = 120.0
ybox = 120.0
zbox = 120.0
boxsigma = 4.0
>>>>

```

*DH* → NLOCAL(1-2/1-2) 7 11

*3SPN.1* → i\_use\_atom\_protein = 0

*Intra mol 1,2* → electrostatic

*Inter mol 1-2* → cutoff\_ele = 20.0

# Simulation of nucleosome



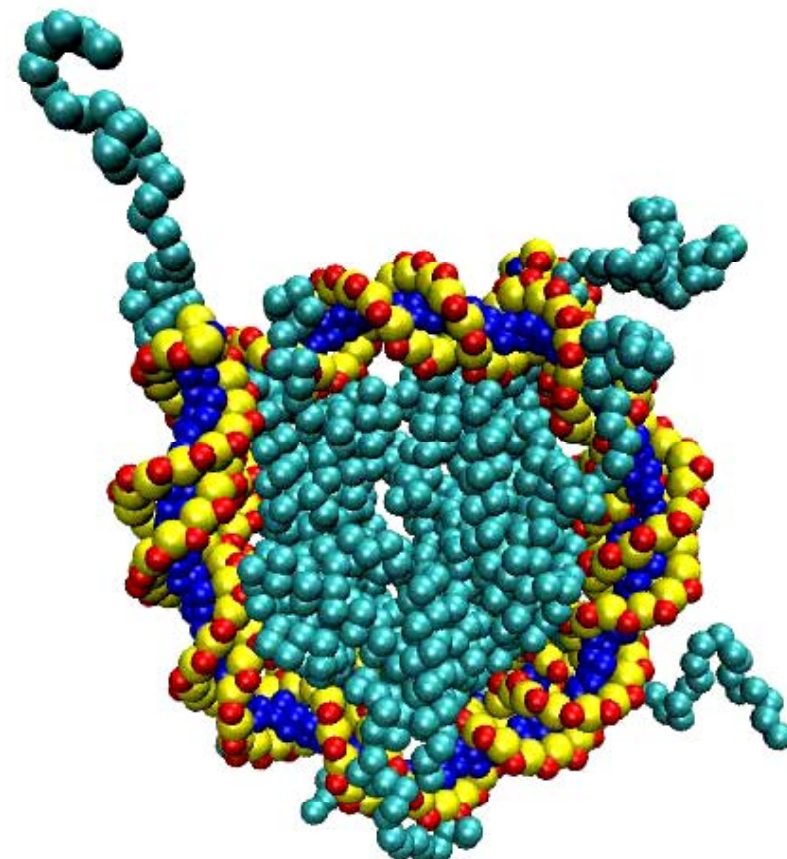
1KX5.pdb

- Electrostatic interaction + Go potential

$$\epsilon_{go}^{pro-dna} = 0.5\epsilon_{go}^{pro}$$
$$[Na^+] = 50mM$$

$\epsilon_{go}^{pro-dna}$ : coefficient of protein-DNA Go potential

```
<<<< energy_function
NLOCAL(1-2/1-2) 7 11
NLOCAL(1-2/3-10) 2 3 7
NLOCAL(3-10/3-10) 2 3
i_use_atom_protein = 0
i_use_atom_dna = 0
>>>>
<<<< electrostatic
cutoff_ele = 20.0
ionic_strength = 0.05
diele_water = 78.0
>>>>
```

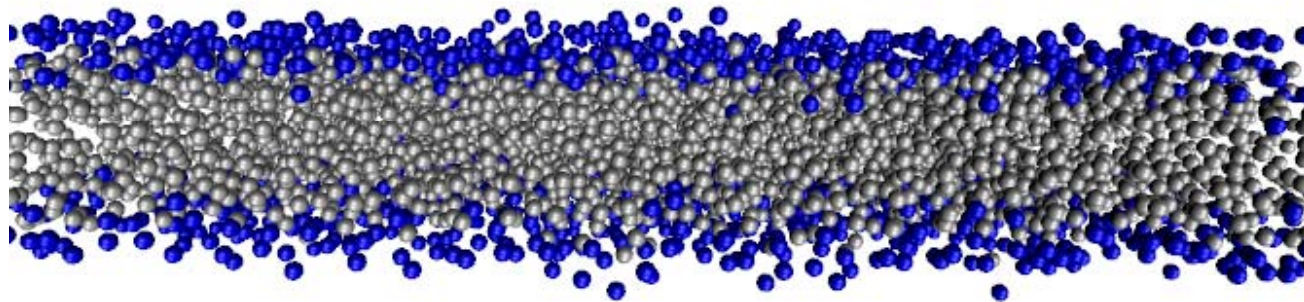


H. Kenzaki, et al unpublished data

# CG lipid model

H. Noguchi, and M. Takasu, Phys. Rev. E (2001)

- $i$  th lipid molecule:
  - 1 hydrophilic particle ( $j=1$ , blue)
  - 2 hydrophobic particles ( $j=2,3$ , gray)
- Self-assemble bilayer structure



# CG lipid model

$$V_{lip} = V_{local} + V_{rep} + V_{hydro}$$

$\theta$ : bond angle

$$V_{local} = K_b \sum_{i,j=1,2} \left( r_{(i,j),(i,j+1)} - r_{0(i,j),(i,j+1)} \right)^2 + K_\theta \sum_i \cos \theta_i$$

$$V_{rep} = \varepsilon \sum_{i \neq i', j}^{N_{st}} \exp \left[ -20 \left\{ \frac{r_{(i,j),(i',j')}}{\sigma} - 1 \right\} \right]$$

$$V_{hydro} = \varepsilon \sum_{i,j=2,3} \begin{cases} -0.5\rho & (\rho_{i,j} < \rho_j^* - 1) \\ 0.25(\rho_{i,j} - \rho_j^*)^2 - c_j & (\rho_j^* - 1 \leq \rho_{i,j} < \rho_j^*) \\ -c_j & (\rho_j^* \leq \rho_{i,j}) \end{cases}$$

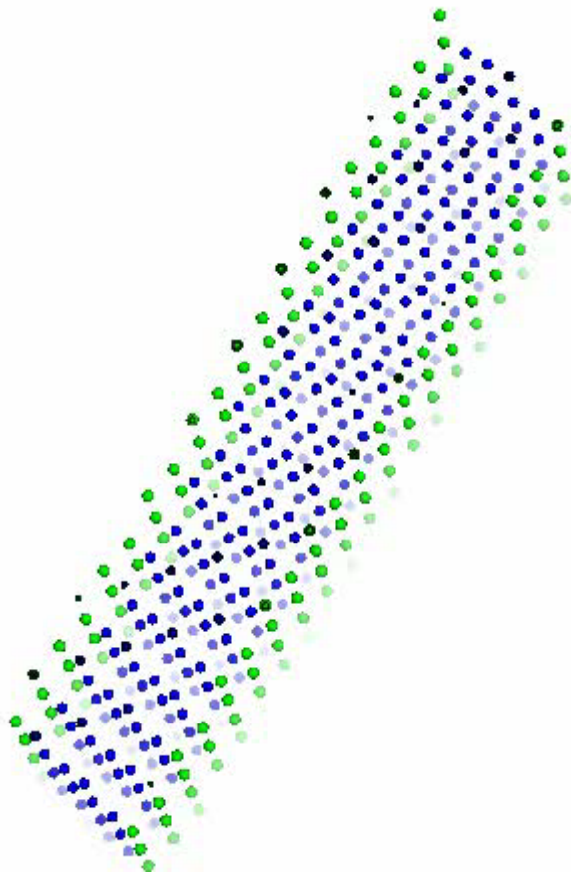
$$\rho_{i,j} = \sum_{i \neq i', j'=2,3} \frac{1}{\exp \left[ 20 \left( \frac{r_{(i,j),(i',j')}}{s} - 1.9 \right) \right] + 1}$$

$$\begin{aligned} K_b &= 500\varepsilon \\ K_\theta &= 500\varepsilon \\ \varepsilon &= 0.6594 \text{ kcal/mol} \\ \sigma &= 7.5 \text{ \AA} \\ \rho_2^* &= 10 \\ c_2 &= 4.75 \\ \rho_3^* &= 14 \\ c_3 &= 6.75 \end{aligned}$$

Density dependent attraction

# Formation of vesicle

S. Fujiwara, et al unpublished data



```
<<<< job_cntl
i_run_mode = 2
i_simulate_type = 1
i_initial_state = 5
>>>>
<<<< unit_and_state
i_seq_read_style = 3
i_go_native_read_style = 3
1 lipid sequence
>>>>
<<<< initial_lipid
nmp_transverse_lipid = 20
nmp_longitudinal_lipid = 20
nlayer_lipid = 1
grid_size_lipid = 1.075
z_coord_lipid(1) = 1.0
>>>>
<<<< energy_function
NLOCAL(1/1) 17 19
i_use_atom_protein = 0
i_use_atom_dna = 0
>>>>
```



# Acknowledgement

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